Heat Shock Proteins and Cardiovascular Disease

Marzie Zilaee^{*}, Gordon A.A. Ferns^{†,1}, Majid Ghayour-Mobarhan^{‡,2}

*Biochemistry of Nutrition Research Center, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

[†]Division of Medical Education, Brighton & Sussex Medical School, University of Brighton, Brighton, United Kingdom

[‡]Cardiovascular Research Centre, School of Medicine, Mashhad University of Medical Sciences,

Mashhad, Iran

¹Equal contribution with first author.

²Corresponding author: e-mail address: ghayourm@mums.ac.ir

Contents

1. Introduction				
1.1 Cardiovascular disease	75			
1.2 Heat shock proteins	78			
Atherogenesis and HSP	83			
2.1 HSPs and CVD in animal models	85			
2.2 HSPs and CVD in human studies	85			
2.3 Molecular mimicry	91			
2.4 HSP and risk factors of atherosclerosis	93			
2.5 The role of HSPs in other common CVDs	95			
Therapeutic Implications	97			
HSPs as Predictors of CVD	98			
Immunization and Vaccination with HSP	99			
Effects of Therapeutic Interventions on HSP	101			
6.1 Herbal medicines	101			
6.2 Statin therapy	101			
Conclusion	101			
References				
	 1.1 Cardiovascular disease 1.2 Heat shock proteins Atherogenesis and HSP 2.1 HSPs and CVD in animal models 2.2 HSPs and CVD in human studies 2.3 Molecular mimicry 2.4 HSP and risk factors of atherosclerosis 2.5 The role of HSPs in other common CVDs Therapeutic Implications HSPs as Predictors of CVD Immunization and Vaccination with HSP Effects of Therapeutic Interventions on HSP 6.1 Herbal medicines 6.2 Statin therapy Conclusion 			

Abstract

Atherosclerosis is the leading global cause of mortality, morbidity, and disability. Heat shock proteins (HSPs) are a highly conserved family of proteins with diverse functions expressed by all cells exposed to environmental stress. Studies have reported that several HSPs may be potential risk markers of atherosclerosis and related cardiovascular diseases, or may be directly involved in the atherogenic process itself. HSPs are expressed by cells in atherosclerotic plaque and anti-HSP has been reported to be

increased in patients with vascular disease. Autoimmune responses may be generated against antigens present within the atherosclerotic plaque, including HSP and may lead to a cycle of ongoing vascular injury. It has been suggested that by inducing a state of tolerance to these antigens, the atherogenic process may be limited and thus provide a potential therapeutic approach. It has been suggested that anti-HSPs are independent predictors of risk of vascular disease. In this review, we summarize the current understanding of HSP in cardiovascular disease and highlight their potential role as diagnostic agents and therapeutic targets.

ABBREVIATIONS

Apaf-1 apoptotic protease activation factor 1 CP Chlamydia pneumoniae CRP C-reactive protein CVD cardiovascular disease DC dendritic cells GGA geranylgeranylacetone hHSP60 human HSP60 HSPB1 HSP-binding factor 1 HSPs heat shock proteins IL interleukin LDL low-density lipoprotein mHSP65 microbial HSP65 MI myocardial infarction SMCs smooth muscle cells TNF tumor necrosis factor

1. INTRODUCTION

Heat shock proteins (HSPs) are highly conserved families of proteins first discovered in 1962 [1]. Recent studies suggest that HSPs and an autoimmune response directed against them may be involved in the pathogenesis of atherosclerosis [2]. Some HSPs are expressed by cells within the atherosclerotic plaque and plasma anti-HSP titers have been reported to be increased in patients with vascular disease. There is a relationship between the serum concentration of some anti-HSP, particularly, anti-HSP27, -60/ 65, and -70 and the manifestations of atherosclerosis and its progression [3]. The strong association between HSP and cardiovascular disease (CVD) suggests that a better understanding of this relationship may allow the development of new therapeutic approaches [4].

1.1. Cardiovascular disease

CVD is a common and major cause of mortality and morbidity [5]. Established risk factors for CVD include age, male gender, family history of CVD, hypertension, hypercholesterolemia, smoking, diabetes mellitus, socioeconomic status, and obesity. Emerging risk factors include oxidative stress, inflammation, and autoimmunity [6].

1.1.1 Atherosclerosis

Atherosclerosis is associated with the accumulation of lipids including modified low-density lipoprotein (LDL) and extracellular matrix in the intima of arteries [7]. Extensive mononuclear cell infiltration and smooth muscle proliferation are characteristics of most atherosclerotic plaques [8]. Atherogenesis is a multistep process that requires an ordered sequence of events in which progress along the disease pathway is driven by risk factors that differ with disease stage [9]. Progression to the fibrofatty plaque is characterized by an enriched content of smooth muscle cells (SMCs) that migrate to the intima from the tunica media. The advanced plaque that forms may subsequently be stable or unstable dependent on its mechanical configuration and cell composition. An unstable plaque is associated with a thin smooth muscle cap and a large lipid pool with a large number of inflammatory cells in the shoulder region [10]. Atherosclerosis starts early in life and slowly progresses for decades. It remains the major cause of most forms of acute coronary syndrome (ACS) including myocardial infarction (MI) and angina, sudden death, and stroke. Inflammation and autoimmunity appear to have a pivotal role in the progression of disease [11] and will be described below.

1.1.1.1 Atherosclerosis and inflammation

The role of inflammation in atherosclerosis was first suggested in the 1850s [12]. More recently immunocytochemical techniques have allowed the cellular composition of atherosclerotic plaques to be determined [13]. These studies have shown that inflammatory cells are present at all stages of development [14]. The earliest lesions in atherogenesis are fatty streaks which have been observed in infants and young children [15]. They are characterized by a relative paucity of lipids and abundance of intimal inflammatory cells including activated T lymphocytes, mast cells, macrophages, and dendritic cells (DC) [16]. Granulocytes and natural killer cells, however, are less common [17].

of numbers Lesions atherosclerosis also contain large of T lymphocytes [18], most of which are T helper type 1 cells bearing α/β receptors [17]. Activated T cells bearing γ/δ receptors are also abundant at the earliest stages of atherogenesis [19] which can be inhibited in experimental animal models by T lymphocyte depletion [20]. The activation of T cells within atherosclerotic plaques indicates that an autoimmune mechanism may be involved. This response is characterized by production of Th1 cytokines and development of a dysfunctional endothelium. The latter occurs as an early event in atherogenesis with subsequent formation of the fatty streak containing lipid-laden foam cells and T cells [21]. Xu et al. reported that CD4+ cells predominate within the T-cell population in early lesions [13], while an increased CD8/CD4 ratio in both early and late lesions was reported by Van der Wal and coworkers [22]. Serum C-reactive protein (CRP) has been reported to be a stronger independent predictor of coronary events compared to LDL or total cholesterol [23–25]. It has been observed that increased soluble intercellular adhesion molecule, a marker of endothelial cell activation, was related to increased coronary risk [26]. Its expression in atherosclerotic lesions was also increased [27]. The activation of complement [28] may have a role in endothelial injury during atherogenesis and may be a consequence of autoimmune responses to modified LDL or denatured HSP [29]. The expression of human lymphocytic antigen class II antigen and several cytokines within atherosclerotic lesions supports the involvement of inflammation in atherosclerosis [18].

Blood-derived monocytes enter the intima of the arterial wall and differentiate into macrophages and DC. Modified lipoproteins are taken up by macrophages that are transformed into lipid-laden foam cells. Monocytederived macrophages appear to be involved at all stages of the disease process [30,31] and DC play a crucial role in directing innate or adaptive immunity against potential autoantigens involved in atherosclerotic disease [32].

Some studies have suggested that serum levels of proinflammatory, T helper-1-related cytokines are positively associated with the severity of atherosclerotic disease, or its risk of development [33,34]. For example, increased serum tumor necrosis factor (TNF)- α and interleukin (IL)-6 were reported to be predictors of coronary and CVD. Increased anti-inflammatory cytokine IL-10 was related to a significantly improved outcome of patients with ACS. It has also been reported that there is an association between serum IL-18 and IL-2 and intima-media (I-M) thickness in

carotid atherosclerosis [35,36]. The relationship between atherosclerosis and inflammation is summarized (Fig. 2.1).

1.1.1.2 The potential role of infection in atherogenesis

The potential contribution of chronic infection to the induction and progression of atherosclerosis and plaque rupture has been controversial [37]. The "infection hypothesis" of atherosclerosis has been investigated by epidemiologic and histopathologic studies, and clinical eradication trials [12]. Animal models and histopathological studies have also addressed the "infection hypothesis" directly. Recent studies on apoE knockout mice have been inconsistent; some studies indicate a potential causal relationship between infection with multiple pathogens and progression of atherosclerosis [38,39] while in another study no relationship between atherosclerosis progression and infection was found [40]. Bacterial DNA of several periodontal pathogens has been found in coronary atherosclerotic plaques, for example, Porphyromonas gingivalis [41]. Chlamydia pneumoniae was also found in the atherosclerotic arterial wall [42,43]; an analysis of 43 histopathological studies has reported that C. pneumoniae was present in 46% of atheromatous arteries and in less than 1% of healthy arteries [44]. A number of large clinical trial including the Weekly Intervention with Zithromax for Atherosclerosis and its Related Disorders (WIZARD), the Azithromycin in Acute Coronary Syndromes (AZACS), the Azithromycin and Coronary Events Study (ACES), and the Pravastatin or Atorvastatin Evaluation and Infection Therapy-Thrombolysis in Myocardial Infarction (PROVE-IT-TIMI22) have investigated the effectiveness of different antibiotics and therapeutic

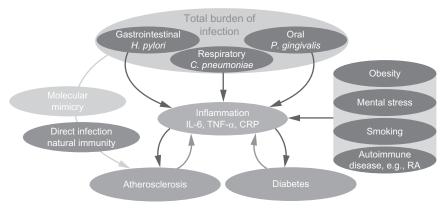


Figure 2.1 Atherosclerosis and inflammation.

regimens on the risk of CVD in patients with ischemic heart disease with or without serological evidence of *C. pneumoniae* infection. The evidence of a short-term reduction in the risk of CVD was weak and there was no evidence of a long-term reduction in these trials [21,45–47]. Antibiotic treatment for *Helicobacter pylori* infection was again inconsistent [48]. A decrease in cardiovascular risk markers has been reported in some studies; for example, plasma cholesterol, LDL cholesterol, fibrinogen, and IL-8; and a reduction of restenosis rates after percutaneous transluminal coronary angioplasty [48–50]. Other studies have reported no significant effect [51]. Pesonen and coworkers [52] reported that the presence of antibodies to several microorganisms was positively associated with carotid intimal thickening, an atherosclerotic marker, in young children. It has been suggested that infection acquired during childhood may lead to atherosclerosis in later life [53]. A positive correlation between CVD extent and infectious organism exposure has been reported [54].

1.1.1.3 Autoimmunity in atherogenesis

It has been widely reported that inflammation and immunity play an important role in the atherosclerotic process [55]. Several potential autoantigens have been identified including modified LDL (oxidized LDL and malondialdehyde-modified LDL) and β -2-glycoprotein I antigen presenting cells such as macrophages and DC present within atherosclerotic lesions, and hence it has been proposed that an autoimmune reaction may be initiated within atherosclerotic plaques [56].

HSPs may be also involved in an autoimmune response predisposing to atherogenesis. Investigation of HSP may provide a further insight into the prevention, prediction, diagnosis, and treatment of CVD.

1.2. Heat shock proteins

HSPs were originally found to be expressed in response to heat, as their name suggests [57]. "Stress proteins" may be more appropriate than "heat shock proteins." However, for historical reasons, the name endures. The discovery of genes expressed in stressful states was first made in fruit fly salivary glands following exposure to extreme heat, that is, chromosome "puffing" [58]. These highly conserved proteins display high sequence homology between prokaryotes and eukaryotes and between different species, a characteristic likely associated with their protective role against stressful environmental conditions [59]. Other stress stimuli include exposure to heavy metals, inflammatory cytokines, amino acid analogues, oxidative

stress, nutritional deficiency, ultraviolet radiation, chemicals, viruses, and ischemia–reperfusion injury [11]. Several risk factors for CVD can also induce HSP, for example, inflammation, ischemia, oxidized LDL, oxidative stress, and hypertension [11].

HSPs have been classified into seven major families on the basis of their molecular weight [3]. These include HSP10 (HSPE), small HSP (15–30 kDa, of which HSP27 (HSPB1) is a member), HSP40 (DNAJ), HSP60 (HSPD), HSP70 (HSPA), HSP90 (HSPC), and HSP100. HSPs have various functions apart from the stress response. Under physiologic conditions, HSPs play an important main role as molecular chaperones by promoting the correct protein folding. Some HSPs are also involved in the transportation of proteins across intracellular membranes and repair of denatured proteins. Although HSPs are protective proteins expressed at high levels in cells under stress, they are also expressed at low concentrations basally [60]. Functions of HSP include the regulation of essential cell functions, such as protein translocation, refolding, assembly and the recognition, and renaturation of misfolded proteins [61,62].

HSPs can therefore prevent the aggregation of denatured proteins, inhibit apoptosis [63], and maintain cellular integrity by stabilization of the cytoskeleton [64]. These functions of HSP are evident in the human arterial wall where HSPs have been shown to be important mediators of protective pathways as well as targets for autoimmunity that may exacerbate atherosclerosis [65,66]. HSPs are normally intracellular proteins, but upon release may induce an autoimmune response [52,67]. Some HSPs have been shown to have immune regulatory responses and modify innate and adaptive immune responses [68] and stimulate the production of proinflammatory cytokines [68].

The cell expression, functions, and reported utility of HSPs as circulating biomarkers are summarized in Table 2.1. Wick *et al.* proposed that the close sequence homology between microbial and human HSP60 (hHSP60) may lead to endothelial cell damage. Early atherosclerosis may involve an immune response mounted against microorganisms that may ultimately lead to an autoimmune response [102]. When HSPs are present on the cell surface or released into the extracellular environment during conditions such as necrotic cell death or viral infection, they may have an immune-stimulatory effect [3]. Surface expression of HSPs as cryptic antigens allows the immune system to recognize them as foreign. Immune responses mounted against microbial HSP may also cross-react with homologous host proteins in a form of molecular mimicry [103]. The autoimmune responses targeted to HSP60

	Other nomenclature	Prokaryotic homologue	Pathological involvement	Cardiovascular expression	Cell functions	Reported potential as biomarkers	
HSP						Antigen	Antibody
HSP 20	HSP B6	_	-	Skeletal muscle and heart muscle [69]	Vasorelaxation [70] Suppresses platelet aggregation [71]	-	-
HSP 27	HSP B [72]	Mycobacterial 18-kDa antigen [73]	-	Smooth muscle cells [74] Endothelial cells [75] Cardiac myocytes [76] Monocytes/ macrophages [77]	 Actin stabilization Muscle contraction Cell migration Cell survival [78] 	Atherosclerosis – Acute coronary syndrome (ACS) [79]	MI in patients with ACS relative to unstable angina – Acute chest pain [80]
HSP 40	-	-	-	_	Chaperone functions [70]	-	-
HSP 60	HSP D [72]	GroEL (<i>E. coli</i>) Mycobacterial 65-kDa antigen [73]	Adjuvant arthritis, rheumatoid arthritis, atherosclerosis, diabetes mellitus, systemic sclerosis, schizophrenia [81]	Ubiquitously expressed [72]	Cell survival [82] Apoptosis [83] – Protein trafficking [84] – Peptide hormone signaling [85]	In carotid atherosclerosis [86] - Associated with IMT in borderline hypertension [87] - Associated with severity of CVD - Infection, stress, myocardial necrosis [88]	 In carotid atherosclerosis Associated with severity of CVD [89] MI compared to CVD Predicative of 5-year mortality in carotid atherosclerosis [90] Higher risk of new CV event [91]

 Table 2.1 HSP: cell expression, intra/extracellular functions, and circulating biomarkers

HSP 70	HSP A [72]	DnaK (<i>E. wlî</i>) [73]	Tuberculosis, leprosy, filariasis, atherosclerosis [81]	Smooth muscle cells [92] Cardiac myocytes [93] Monocytes/ macrophages [92]	Anti-inflammatory [92] – Antiapoptotic – Antioxidant [94]	Levels associated with decreased IMT in hypertensive patients [95] - Levels associated with low CVD risk - Carotid atherosclerosis - Inversely correlated with neutrophil activation - ACS [96]	CVD [97] – Not related with prevalence of CVD [95] and high risk of ACS [98]
HSP 90	HSP C [72]	C62.5 (E. coli) [73]	Schistosomiasis, systemic lupus, erythematosus [81]	Macrophages [92] – Smooth muscle cells [92]	Antioxidant – Antiapoptotic – Proangiogenic [99]	Atherosclerosis [100]	Atherosclerosis [101]

appear a consistent feature of atherosclerosis suggesting that this HSPs may play a role in the pathogenesis of atherosclerosis [104]. The expression of some HSPs is increased in regions of the artery wall containing atheromatous plaques [81,105]. Following cerebral ischemia, protein synthesis in the brain is generally suppressed. However, some specific genes are expressed in increased levels, and these include the genes for HSPs and amyloid precursor protein [106].

Most HSPs facilitate the correct folding of denatured proteins. These complex interactions between chaperones, co-chaperones, and their client proteins lead to either a new folding attempt or protein ubiquitination and subsequent degradation via the proteasome pathway [107].

Some HSPs can inhibit apoptosis [107]. Apoptosis involves the release of cytochrome *c* from mitochondria. It is binding to apoptotic protease activation factor 1 (Apaf-1) triggers oligomerization. This complex then attracts the inactive unprocessed pro-form of the proteolytic enzyme caspase-9, which is cleaved to its active form, thereby initiating the apoptotic process. HSP90 binds Apaf-1 thereby preventing its binding to cytochrome *c*, whereas HSP70 prevents oligomerized Apaf-1 from recruiting pro-caspase-9 [107].

HSP90 has also been shown to bind to endothelial nitric oxide synthetase in the heart and increase its activity [108]. With increasing age, the induction of HSP by stressful stimuli appears to diminish in a variety of tissues including the heart due to impaired activation of heat shock transcription factor 1 (HSF-1) by stress [108]. This phenomenon was associated with a reduction in the protective effect of mild heat shock or ischemia against subsequent severe ischemic stress in the aged hearts cells in animal studies [108,109].

Individuals with chronic infection have high serum HSP60. These individuals also tend to have poorer diet and living conditions that could potentially confound this observation [47]. A positive relationship between the immune responses to HSP following immunization with bacillus Calmette–Guerin (BCG), an attenuated strain of tuberculosis, and subsequent atherosclerosis has been reported in a rabbit model of atherosclerosis [110]. This finding suggested that exposure to specific microorganisms may promote atherogenesis.

1.2.1 Production of HSPs

Upregulation of HSPs is mediated by the release of HSF1 in response to stressful stimuli such as increased temperature [111] or ischemia [112].

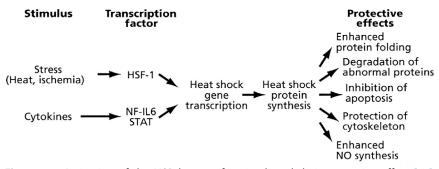


Figure 2.2 Activation of the HSP by specific stimuli and their protective effect [73]. Chronic inflammatory conditions such as smoking, stress, obesity, and rheumatoid arthritis may contribute to the total burden of inflammation and hence to atherosclerosis. *Ref.* [89]. *From P.J. Ford, G.J. Seymourm, K. Yamazaki, Cardiovascular and oral disease interactions: what is the evidence? Prim. Dent. Care* 14(2) (2007) 59–66.

Monomeric cytoplasmic HSF1 forms a trimer and translocates to the nucleus where it binds to its target sites (heat-shock elements) in the regulatory region of the HSP genes (Fig. 2.2) [30]. The activity of HSF1 is negatively regulated by the binding of HSP70 to its transactivation domain and the resultant repression of heat shock gene transcription [100]. There is also an interaction between HSP-binding factor 1 (HSPB1) with the active trimeric HSF1 and HSP70 thereby inhibiting the capacity of HSF-1 to bind DNA [113].

1.2.2 HSP in the extracellular space

The presence of increased plasma HSP concentrations is usually associated with stress including inflammation, and bacterial, and viral infection [114]. HSP exit the cell and enter the extracellular milieu by two mechanisms: passive release usually followed cell damage and active release involving exosomes [115,116] or lysozome-like vesicles [117]. Exosomes are small membrane vesicles that numerous cell types secrete. These contribute to HSP70 release from human peripheral blood mononuclear cells in both basal and stress-induced states [115]. Infections may represent a nonendogenous source of HSP in the extracellular compartment [118,119].

2. ATHEROGENESIS AND HSP

The importance of HSPs in atherogenesis is unclear. HSP expression and reactivity thereof appear to be associated with the inflammatory response that is characteristic of atherogenesis [120]. The intensity of HSP expression is positively associated with atherosclerotic severity. Immunization with recombinant mycobacterial HSP65-induced atherosclerotic lesions in normocholesterolemic rabbits [121], normal C57BL/6J mice fed a high-fat diet [122], and LDL receptor-deficient mice [123]. In early atherosclerotic lesions, only DC were found to express HSPs [124]. Furthermore, in advanced atherosclerotic lesions, HSPs were found to be expressed by several cell types including SMC, DC, and monocytes/macrophages.

Inflammation and immunologic reaction, including the autoimmune response against several HSPs, appear to play an important role in atherosclerotic pathogenesis [125]. Both HSPs and anti-HSPs have been shown to stimulate the production of proinflammatory cytokines [48].

Wick and colleagues [124] have hypothesized that an immune response to HSPs, either endogenously derived from cells involved in atherogenesis or exogenously from microorganisms, may lead to complement-mediated endothelial injury and subsequent atherosclerosis. Mayr *et al.* reported that serum anti-HSP directed against *Escherichia coli* and *C. pneumoniae*-mediated lysis of stressed, but not unstressed endothelial cells [126]. Alternately, these immune responses may be induced following exposure of HSPs on the surface of infected endothelial cells [11]. SMC can be induced to express HSPs as part of a survival mechanism following exposure to a variety of stressors, that is, high blood pressure [127].

HSPs are immunodominant molecules. A significant element of the immune response to pathogenic microorganisms is directed toward HSP-derived peptides. HSPs are highly conserved molecules with phylogenetic similarity between microbial and mammalian forms (50–60% identical residues for HSP60). This finding has led to a debate as to whether HSPs might act as potentially harmful autoantigens [128]. Immunologic recognition of cross-reactive HSP epitopes might provide a link between infection and autoimmunity [129]. This premise has been supported by studies implicating immunity to HSPs in arthritis [130,131], multiple sclerosis [132,133], and diabetes [134,135].

Some studies have shown that prior induction of the HSPs by a mild stress has a protective effect against more severe stress. Increased expression of HSPs in cardiac cells in culture or intact heart produced a protective effect similar to antioxidant enzymes [73].

Most previous studies of HSP and CVD have focused on HSP60, 65, and 70 [11]. However, there has been recent interest in HSP27.

2.1. HSPs and CVD in animal models

Reduced atherosclerotic lesion size after subcutaneous immunization with mycobacterial HSP65 has been demonstrated in apoE knockout mice [136]. HSPs released into the circulation from the infarcted heart tissue bind to circulating anti-HSPs. This antigen–antibody complex may then subsequently be removed by the reticuloendothelial system [120].

Reactivity to conserved HSP60 was reported to induce a regulatory T-cell phenotype in animal models of arthritis [137,138]. These findings suggested that altered immune response in the atherosclerotic lesion may influence disease pathogenesis. In fact, a reduction of Th1 polarization of CD4 – T cells in apoE knockout mice by pentoxifylline appeared quite effective in reducing atherosclerosis [139]. The data support the possibility that HSP induced in the early stages of atherogenesis might promote the nonspecific inflammatory response in the vessel wall and the recruitment of monocytes and T cells into the developing inflammatory lesion [140]. The induction of HSP70 expression in rat aortic tissue by heat and stannous chloride treatment was associated with an anti-inflammatory state [141]. The roles of HSP in animal model studies are summarized in Table 2.2.

2.2. HSPs and CVD in human studies

2.2.1 Atherogenesis and HSP20

Kozawa *et al.* reported that serum HSP20 (HSPB6) was increased in cardiomyopathy in hamsters, though its tissue of origin was unidentified. Myocardial angiogenesis was enhanced in hearts that overexpressed HSP20 [71]. HSP20 appeared to act as a cardiokine, regulating myocardial angiogenesis through activation of the VEGFR signaling cascade. HSP20 has also been reported to affect platelet aggregation [147]. HSP20 was found to inhibit thrombin-induced calcium influx without affecting release from calcium stores. Also, HSP20 inhibited the increase in cytoplasmic free calcium triggered by collagen, but not that induced by the calcium ionophoreA-23187 in platelets *in vitro* [148].

The maintenance of redox status is a constant challenge in normal hearts, but even more so under pathological conditions. Small HSPs, in particular, may play a protective role against redox imbalance, that is, when excess production of reactive oxygen species exceeds scavenging mechanisms [149].

Zhang *et al.* observed that circulating HSP20 was increased in a transgenic mouse model with cardiac-specific overexpression [150]. HSP20 was secreted via exosomes independent of the endoplasmic reticulum–Golgi

Study	Subjects	Finding	References
ApoE knockout mice were infected with MCMV and CP	ApoE knockout mice	Infection with MCMV alone, CP alone, and both MCMV and CP increased lesion size of atherosclerosis	[38]
Effect of HSP70 administration on the size of infract	Rat and rabbit	HSP 70 administration limited infarct size following the exposure of the heart to ischemia–reperfusion injury	[142]
Immunization with HSP65 or mycobacterial HSP65	Wild-type C57BL/6J mice	Enhanced the lesion formation	[143]
Effects of antibodies directed against, and lymphocytes reactive to HSP65 on fatty-streak formation	LDL-RD mice	Promote fatty-streak formation	[144]
Immunization with HSP65	Normocholesterolemic rabbits	Promotes atherosclerotic lesion formation	[121]
Immunization with the bacillus Calmette–Guerin (BCG)	Cholesterol-fed rabbit	Enhance atherogenesis	[145]
Effect of high- cholesterol diet on anti-HSP and von Willbrand factor (vWF)	Rabbit	Induce the expression of anti-HSP60, 65, and 70 and thus increasing concentrations of von Willbrand factor (vWF), a marker of endothelial injury	[146]
Effect of Rabbit immunization with HSP60 on T lymphocytes		Depletion of peripheral blood T lymphocytes	[20]

Table 2.2Immunomodularity effects of HSP in cardiovascular disease in animal modelsStudySubjectsFindingReferences

pathway. HSP20 was involved in regulating myocardial angiogenesis via VEGFR signaling cascade activation.

HSP20 expression and its phosphorylation at its serine16 residue was increased following experimental MI. Mouse models of cardiac-specific overexpression of HSP20 subjected to ischemic/reperfusion showed smaller MI and improved recovery of contractile performance during the reperfusion phase [151]. This observation is likely to be due to the ability of HSP20 to protect against cardiomyocyte necrosis and apoptosis. As such, HSP20 may be a new therapeutic target for the prevention and treatment of myocardial remodeling and ischemic injury.

Li *et al.* reported on a proteomic study of cardiac ventricle isolated from failing and nonfailing human hearts [152]. Altered proteins included those associated with metabolic, cytoskeletal, and stress response function. Of the latter, HSP27 and 20 were identified.

HSP20 and its phosphorylation have been implicated in multiple physiological and pathophysiological processes including smooth muscle relaxation, platelet aggregation, exercise training, MI, and insulin resistance. HSP20 has an important role in contractile function and cardioprotection [153].

It has recently been shown that HSP20 triggers apoptosis by interacting with key proapoptotic signaling proteins [154], and its overexpression was shown to protect cultured rat cardiac myocytes from β -agonist-induced apoptosis [155].

Qian *et al.* have generated a cardiac-specific overexpression model in which HSP20 serine 16 was substituted with alanine [156]. This model was subjected to ischemia/reperfusion to ascertain if the cardioprotective effects of HSP20 were associated with serine16 phosphorylation. Interestingly, prevention of phosphorylation influenced the balance between cell death and autophagy. The HSP20 aggregation pattern was also altered. Prevention of phosphorylation attenuated cardioprotection against ischemia/reperfusion injury via suppressed autophagy and increased cell death.

HSP20 has been shown to enhance myocardial contractile function and protect against doxorubicin-induced cardiotoxicity [157]. Wang *et al.* investigated the role of HSP20 in sepsis-mediated cardiac injury by examining the expression profiles of five major HSP in response to lipopolysaccharide challenge [158]. HSP20 expression was downregulated in lipopolysaccharidetreated myocardium suggesting that this decrease might contribute to cardiovascular defects. In contrast, overexpression of HSP20 significantly enhanced cardiomyocyte contractility following lipopolysaccharide treatment. This study indicated that HSP20 overexpression prevented endotoxin-induced myocardial dysfunction and apoptosis via inhibition of NF- κ B activation.

 β -Adrenergic neurohormonal axis stimulation contributes to the progression of heart failure and mortality in animal models and patients. Activation of the β -adrenergic pathway in cardiomyocytes has been shown to result in transiently increased cardiac HSP20 expression [159]. Cardiac HSP20 overexpression may protect the heart against β -agonist-induced cardiac remodeling. HSP20 attenuated the cardiac hypertrophic response, markedly reduced interstitial fibrosis and decreased apoptosis.

2.2.2 Atherogenesis and HSP27

HSP27 is a 27-kDa protein ubiquitously expressed by many cell types including vascular cells. HSP27 is expressed at high levels in a variety of tumors and normal tissues including the heart [160]. HSP27 has several roles, including regulating apoptosis and is involved in vascular SMC migration and proliferation, embryogenesis, cardioprotection, resistance to oxidative stress, and modulation of inflammation [161,162]. These roles may have an important bearing on atherogenesis. HSP27 may have cardioprotective effects by mechanisms that include increasing myocardial resistance to oxidative stress and its effects on apoptosis [163]. Some studies have indicated that HSP27 overexpression protected cardiac myocytes against ischemic injury [161,164] and promoted cell protection and survival during inflammation by regulating anti-inflammatory gene expression [165]. Reports have suggested that HSP27 was associated with acute cardiac chest pain [166] and ACS [167]. Increased HSP27 expression was related to improved right ventricle function and systemic perfusion [168]. Park et al. found that serum HSP27 was increased in the early hours following ACS, but fell to near normal levels about 12 h from chest pain onset [169].

HSP27 may be a putative autoantigen involved in atherogenesis [170]. It was reported that high antibody titers against HSP27 were associated with cardiovascular events [171]. Although HSP27 IgG was significantly increased in patients with chest pain versus healthy control subjects, antibody levels were similar in both troponin I positive (MI) and negative patients [167].

Anti-HSP27 has been detected in plasma of patients with CVD. Although increased serum anti-HSP27 has been found in patients with AMI, the evidence for its association with CVD has not been fully established [80]. Shams *et al.* found increased anti-HSP27 in patients with chest pain [153]. Anti-HSP27 was increased during the first 12 h following an ACS event, then fell to near normal after 12 h. Ghayour-Mobarhan *et al.* reported that serum antibody titers of HSP27 increased and decreased rapidly after onset of ACS [154], suggesting that anti-HSP27 may be an early marker of MI and unstable angina.

2.2.3 Atherogenesis and HSP60/65

The HSP60 family of proteins is involved in development of several diseases including arthritis in rats, rheumatoid arthritis in humans, insulin-dependent diabetes mellitus in mice, and systemic sclerosis in humans [172]. Several studies have reported that increased anti-HSP60 was associated with CVD. Recent studies indicate that high titers of anti-HSP60 were associated with coronary atherosclerosis [89,173]. Children with increased anti-HSP60 had an \sim 10-fold chance of being from a high-risk CVD family versus children with normal anti-HSP60 concentration [174]. Anti-HSP may initiate complement activation [175] thereby increasing vessel wall inflammation, that is, a process that plays an essential role in atherosclerosis [91,176]. Some studies suggested that plasma HSP60 was increased early in heart failure [177].

Significantly increased serum anti-HSP65 was observed in patients with CVD [90] and carotid atherosclerosis [178]. A strong correlation was observed between increased anti-HSP65 and restenosis after percutaneous transluminal coronary angioplasty [179]. In patients with CVD, anti-HSP60 and anti-HSP65 were significantly associated with the presence and severity of disease. The concentration of HSP60 and 65 has been related to more severe CVD forms [180]. Some conditions, such as cardiac failure, result from heart damage caused by a variety of factors [181]. Heart damage due to stress factors can lead to sudden death because of cardiac dysfunction [182]. Serum anti-HSP60/65 concentration was associated with CVD severity and progression [89,98]. One study investigated the effect of cardiac rehabilitation therapy, statin treatment, or combination of both on anti-HSP in patients with CVD after percutaneous coronary intervention [178]. This study showed that reduced anti-HSP was accompanied by greater improvement in blood biochemical variables including lipids, high-sensitivity CRP (hsCRP), and IL-6. HSP and anti-HSP elicited the production of proinflammatory cytokines by macrophages and adhesion molecules [183]. There was no significant relationship between antibody titers to HSP and other inflammatory factors (hsCRP and IL-6), suggesting they are independent risk factors for CVD [184]. HSP60 expression was localized to atherosclerotic lesions as opposed to nonatherosclerotic regions of the arterial wall [185].

In animal studies, anti-HSP60, 65, and 70 were correlated to the extent of atherosclerosis as well as markers of endothelial dysfunction [146]. Furthermore, HSPs were potential targets for the immune response, may be altered during the stress response, and thereby directly contribute to the inflammatory process [11].

These immune responses may initially be directed against antigens present on pathogenic organisms and then cross-react with homologous host HSP elaborated by cells of the vascular wall including endothelial cells [186]. It has been reported that a cellular and humoral response to HSP65 was present in subjects with carotid and coronary atherosclerosis [90,187].

Kleindienst *et al.* demonstrated that HSP60 was present on endothelial, smooth muscle, and mononuclear cells of carotid and aortic specimens [17]. Increased plasma anti-HSP was associated with the presence and progression of vascular disease. Increased antibody against mycobacterial HSP65 was reported in patients with carotid atherosclerosis [187], coronary heart disease [90], and borderline hypertension [188]. Serum anti-HSP60 was increased in subjects with peripheral vascular disease [150]. Anti-HSP65 might be prognostic because its concentration appeared to predict 5-year mortality in patients with carotid atherosclerosis [98]. Wysocki *et al.* showed that anti-HSP60 was significantly higher in ACS versus controls [151]. Giuseppina *et al.* showed that HSP60 may be an independent predictor of post-MI adverse cardiovascular events [152].

2.2.4 Atherogenesis and HSP70

HSP70 (prokaryotic homologue: DnaK (*E. coli*)) is involved in protein folding [3]. In the rat, serum HSP70 was related to the extent of experimentally induced MI [155]. Larger infarct after focal ischemia and more abundant apoptotic cell death after transient focal ischemia were associated with lower HSP expression. Previous studies have shown a strong positive correlation between plasma anti-HSP60, -65, -70, and -72 with CVD [167]. HSP70 expression was associated with the extent (thickness) of the atherosclerotic plaque. HSP70 distribution in the human aorta showed a homogeneous staining pattern in "normal-appearing" regions, but a heterogeneous pattern in areas of atherosclerosis. It was reported that the heterogeneous pattern of HSP70 distribution in atherosclerotic lesions may be due to the leakage of HSP70 from damaged cells into the plaque [126]. Synthesis of HSP normally protects cells from death, but it has been hypothesized that insufficient HSP70 in SMC of the aorta leads to their death and further promotes plaque rupture and thromboembolic complications [189]. Although HSP70 expression was increased in advanced atherosclerotic lesions and several cell types (monocytes, macrophages, DC, and SMC), only DC expressed high levels of HSP70 in early lesions [70].

Pockley *et al.* reported that HSP70 expression was localized to the center of thickened atheromatous plaques and that staining intensity correlated with atherosclerotic plaque thickness [95]. Several cross-sectional studies [95,156] have reported that HSP70 may have an atheroprotective role, a finding that may be mediated by its effect on SMC survival. It was subsequently shown that location of HSP70 expression changed during plaque evolution and was positively associated with severity of atherosclerosis and altered protein distribution [158].

Although increased serum HSP70 was associated with a lower risk of CVD independent of established CVD risk factors, some induced HSP70 expression by endothelial cells and SMC. Zhu *et al.* have also reported that the severity of coronary disease (number of diseased vessels) was inversely related to serum HSP70 [157]. Increased HSP70 was reported in patients with chronic heart failure [159]. Because HSP represented vessel wall cellular response to stress-inducing factors, including several classical atherosclerosis risk factors, these may have roles in atherogenesis [190]. Interestingly, increased HSP70 was associated with low CVD risk, suggesting a complex role for these proteins in atherosclerosis [180].

2.3. Molecular mimicry

The potential for cross-reactivity of the immune response to bacterial HSP with hHSP60 expressed on stressed endothelial cells has been proposed as a possible mechanism for the early atherogenic involvement of HSP60/65 in atherogenesis. Infections have been implicated in CVD [136]. The fact that microorganisms produce HSP homologous to human HSP has given rise to the concept of molecular mimicry [61]. A high degree of sequence identity between human and microorganism HSP is one potential mechanism to explain its association with infection, high HSP expression, autoimmunity, and CVD [126]. GroEL (a bacterial HSP) is highly immunogenic [128]. It has been proposed that GroEL immune response cross-reacted with hHSP60 expressed on endothelial cells leading to endothelial dysfunction and atherogenesis [126]. The immune system may not be able to differentiate HSP of human and bacteria origin and thus molecular mimicry may provide a mechanism for autoimmunity (Fig. 2.3).

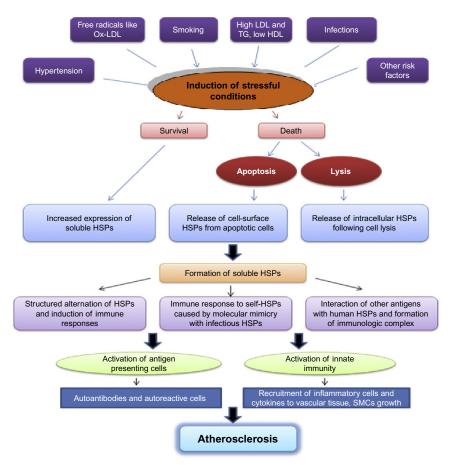


Figure 2.3 Possible mechanisms of involvement of HSP in atherosclerosis.

It has been reported that cellular immunity against HSP60 is related to I-M thickness in young male individuals but not in the elderly, suggesting a possible role of specific cellular immunity to HSP60 in early stages of atherosclerosis [191]. However, these results were not consistent with those of the Bruneck study [192] which showed no relationship between circulating HSP60-specific T cells and late stages of atherosclerosis. Moreover, Ramage *et al.* have reported that the proliferative response of human T lymphocytes to highly purified hHSP60 was confined to the adult CD45RA – RO + naïve subset, whereas both memory and naïve T-cell populations proliferated in response to bacterial HSP60 [193]. In patients with different auto-immune conditions and those with established atherosclerosis, increased

T cell response to microbial HSP65 (mHSP65) as well as increased circulating anti-mHSP65 and anti-HSP60 have been reported [194]. These immune responses may initially be directed against antigens of pathogenic organisms and then cross-react with homologous host HSP elaborated by vascular wall cells, including endothelial cells [186]. Anti-HSP may lead to endothelial injury by antibody-dependent complement-mediated cellular cytotoxicity, whereas anti-HSP, probably produced for the primary purpose of eliminating infectious organisms, may lead to endothelial injury. Mayr et al. have reported that serum anti-HSP to E. coli and C. pneumoniae can mediate endothelial cell lysis of stressed, but not unstressed endothelial cells [126]. Alternatively, these immune responses may be induced following exposure of HSP on the surface of infected endothelial cells [61]. In atherogenesis, SMC have an important role and can be induced to express HSP as part of a survival mechanism following exposure to a variety of stressors including high blood pressure. HSP have an important role in the function of cells involved in the immune system. Because of their distribution and homologous structure in different species, they may serve as autoantigens. A positive relationship of HSP immune response with atherosclerosis has been observed in rabbit models [110]. The possible mechanisms by which HSP may be involved in atherosclerosis are summarized in the figure (Fig. 2.3). Anti-HSP60 and 65 were positively related to cardiovascular morbidity and mortality [136]. Lenzi and coworkers reported that increased anti-HSP60 may constitute a marker of CVD [12]. Studies have shown that CVD risk in patients with a systemic immune response was associated with increased HSP [195].

2.4. HSP and risk factors of atherosclerosis

Although it was hypothesized that increased anti-HSP developed relatively early in the atherogenic process, a prospective study would be required to confirm this. However, increased anti-HSP appears associated with CVD risk factors [196].

In a large community-based study, increased soluble HSP60 correlated with LDL cholesterol [197]. Anti-HSP60 was positively associated with indices of obesity, more so than anti-HSP65 and 70 [58]. A positive association was found between serum HSP60 and various psychosocial measures including low socioeconomic status, social isolation, and psychological distress [198].

HSP27 and its antibody concentration were related to cardiovascular complications in patients with glucose intolerance [199]. It was reported that

anti-HSP27 IgG was strongly associated with age, gender, and hypertension, but weakly with diabetics with ACS [196]. Increased anti-HSP70 was also reported in hypertensive men versus normotensive controls [200].

Figueredo *et al.* measured class-specific antibodies to mycobacterial HSP70 in patients with diabetes mellitus with or without vascular complications [201]. IgG and IgM in diabetic patients did not significantly differ from healthy control subjects or between types of diabetes, regardless of gender, disease duration, hemoglobin A1c, or type of vascular complication. Mean serum IgA in type 2 diabetics was, however, significantly higher than matched controls. This result suggested a possible role of anti-HSP70 IgA in the pathogenesis of the vascular complications in diabetes mellitus [202].

Pockley *et al.* determined HSP and anti-HSP in subjects with established hypertension [203]. HSP60 and 70 antigen and anti-HSP60 were similar in hypertensive and normotensive controls, whereas anti-HSP70 and anti-HSP65 were increased. HSP60 was not associated with atherosclerosis. Anti-HSP70 and anti-HSP65 were both associated with hypertension, independent of age, smoking status, and blood lipids.

Kervinen *et al.* studied middle-aged men with MI or coronary death during an 8.5-year follow-up [204]. Control individuals were subjects without coronary events and matched for drug treatment and geographic region. Although anti-HSP60 IgA predicted coronary risk, the effect was modest without simultaneous occurrence of other classical risk factors.

The relationship between HSP and risk factors of CVD is summarized (Table 2.3).

Subjects	Findings	References
Subjects with dyslipidemia	Anti-HSP60, 65, and 70 are significantly increased in dyslipidemic patients	[205]
Diabetic patients	Positive relationship between anti-HSP70 and type-2 diabetes	[202]
Hypertensive patients	Positive relationship between anti-HSP65 and 70 and hypertension	[203]
Patients with CVD	Positive relationship between anti-HSP27 and total cholesterol	[169]
Dyslipidemic middle-aged men	Negative relationship between anti-HSP60 and HDL cholesterol	[206]

 Table 2.3
 HSP and risk factors of CVD

2.5. The role of HSPs in other common CVDs

2.5.1 Cardiac hypertrophy

During the development of cardiac hypertrophy, cardiac chaperones such as HSP70 and 22 are found to have increased expression. Angiotensin II infusion, isoproterenol infusion, and swimming were identified as hypertrophic stimuli of HSP70 in an animal model study [207]. Cardiac hypertrophy is a response to a number of stress stimuli. In cardiac hypertrophy, a general increase in cardiac chaperone expression was not surprising, but the reasons for this finding are poorly understood [208].

It appears that HSP70 is necessary for the induction of cardiac hypertrophy. Overexpression of a dominant-negative form of HSP70 or decreased histone deacetylase 2 with small interfering RNA blunted the hypertrophic response in the heart [207]. In mice, cardiac hypertrophy induced by isoproterenol infusion or aortic banding resulted in a blunted histone deacetylase 2 activity suggesting a role for HSP70 in the induction of cardiac hypertrophy possibly by stabilization of histone deacetylase 2.

The histone deacetylases (HDACs) are key enzymes in the regulation of hypertrophic gene expression [209]. Class II HDAC (HDAC4, HDAC6, HDAC7, and HDAC9) appear to negatively regulate hypertrophy by repressing myocyte enhancer factor/GATA/NFAT-mediated gene transcription [210].

During development of cardiac hypertrophy, HSP20 was increased, and this may lead to activation of signaling pathways involved in survival and cell growth, including PI3K/Akt, AMP kinase, protein kinase C, nitric oxide, and mTOR [201]. These appear associated with several cardioprotective mechanisms including preconditioning, growth, and protection against apoptosis [201].

In a variety of animal models, HSP22 was increased during development of cardiac hypertrophy and in cultured cardiomyocytes and intact mouse heart [211,212]. Increased HSP22 expression resulted in the development of a spontaneous hypertrophy characterized by reexpression of the fetal gene program.

2.5.2 Heart failure

Knowlton *et al.* investigated HSP90, 72, 70, 27, and 60 expression in cardiomyopathy and ischemic cardiomyopathy patients [213]. HSP72, 70, and 90 were unchanged versus normal controls. A twofold increase in heart HSP27 expression was, however, noted in cardiomyopathy. HSP60 contains NF- κ B binding elements. HSP72 does not. This may explain why HSP60 expression was increased in heart failure, whereas other HSP were not [214].

Using a rat model of heart failure, increased HSP72 and 27 expression was noted [215]. HSP60 expression was not changed.

Using the same model, Tanonaka *et al.* reported that increased myocardial HSP60 expression was associated with subsequent development of heart failure [204].

The presence of HSP60 in the blood of normal individuals was first reported in 1999 [216]. HSP60 distribution in the myocardial cells changes in dilated cardiomyopathy and ischemic heart disease by translocation from the cytosol to the mitochondria [217].

In explanted failing hearts, HSP60 was localized to the plasma membrane where it was associated with increased apoptosis [110]. Its interaction with other cells may trigger the innate immune response resulting in release of proinflammatory cytokines such as TNF. This finding may imply that HSP60 is an early effect or molecule that induces myocyte loss and contributes to heart failure. The role of circulating HSP60 in the pathophysiology of heart failure has not been clearly delineated and more research is needed [218].

It has been hypothesized that circulating HSP60 induces inflammation and autoimmune responses that play a role in atherosclerosis [219]. Niizeki *et al.* showed that serum HSP60 was increased in patients with congestive heart failure [220].

2.5.3 Ischemic cardiac injury

A cardioprotective effect for HSPs in ischemia/reperfusion injury has been reported. Ischemia/reperfusion injury resulted in increased HSP70 and HSP90 mRNA [221]. The increase in HSP70 was substantially higher than HSP90. Increased expression was most likely due to a concurrent increase in HSF1 driven by an accumulation of reactive oxygen species [211].

Depre *et al.* investigated the differential expression of genes following ischemia/reperfusion injury in a pig model and found that HSP22 increased significantly after 1 h of reperfusion [212].

In animal studies, increased HSP70 and 72 expression was beneficial following cardiac ischemia/reperfusion injury [222]. HSP72, induced by successive bouts of endurance exercise, led to improved ischemia/reperfusion injury outcome as demonstrated by decreased infarct size and cardiac apoptosis. In mouse models, cardiac-specific HSP20 overexpression of HSP20 protected against ischemia/reperfusion injury [70]. Improved contractile performance, decreased myocyte cell death, and significantly decreased infarct size were noted.

One of the mechanisms for the protective effect of HSP20 appears to be in activating autophagy, a mechanism critical for ischemia/reperfusion injury [213]. Interestingly, transgenic mice in which serine 16 on HSP20 is mutated, that is, nonphosphorylatable, were more susceptible to ischemia/reperfusion injury than wild-type mice.

3. THERAPEUTIC IMPLICATIONS

HSP may act as an autoantigen that results in vascular injury cycle. As such, tolerization may limit atherogenesis [61]. Bacterial HSP60 is immunologically cross-reactive with its mammalian counterparts. The finding that immune responses to HSP are associated with both experimentally induced and spontaneous autoimmune diseases in animals has prompted intense research to assess the role of bacterial HSP as the etiological agents involved in the development of autoimmune disease [223].

Mucosal administration of HSP has been used to induce a state of immune tolerance. Harats and colleagues have shown that HSP65 tolerization led to reduced plaque formation in a murine model of atherosclerosis. Tolerization was also associated with reduced macrophage and T-cell infiltration and increased expression of the anti-inflammatory cytokine, IL-10 [215]. Some HSPs have been used as carriers for delivering epitopes to the immune system in the absence of adjuvant [224]. Whole pathogen vaccines such as BCG contains a high concentration of potentially immunogenic HSP. While reducing morbidity and mortality, these vaccines may also stimulate proatherogenic mechanisms [225]. Indeed, immunization with BCG vaccine increased atherosclerosis in the cholesterol-fed rabbit. Anti-HSP60 was related to plaque formation suggesting that the specific immune response to BCG-associated HSP might be proatherogenic [110,145]. In atherosclerotic lesions induced by BCG immunization alone in the absence of traditional risk factors, the inflammatory response to HSP60 was insufficient to drive atherogenesis over prolonged period of time [226]. HSP27 may also be a potential therapeutic target. Wolfgang et al. suggested that HSP27 had a protective effect against simulated ischemia. They proposed a model in which proteins, not in their final folding state, bind to oligomeric HSP complexes that act as a shelter. After resolution

of ischemia, these proteins are released in their final folded state thereby assuming normal activity in cells recovering from ischemic injury [227]. It has been suggested that serum HSP27 may be a predictive marker of atherogenes which helps to identify individuals who may benefit from estrogen therapy [221]. Targeted HSP27 overexpression in tissues with ischemic injury may also be beneficial. The use of pharmacologic inducers or enhancers of the endogenous HSP expression in the target cells can be used. For example, herbimycin A and geranylgeranylacetone (GGA), a cyclic polyisoprenoid, induced an acytoprotective potential in simulated ischemia studies. Oral administration of GGA rapidly upregulated HSP expression in response to a variety of stressors, while its effect was attenuated under non-stress conditions [228]. Oxidative stress, inflammation, and apoptosis were involved in the initiation, development, and rupture of atherosclerotic plaques. The role of HSP in treatment of CVD continues to receive much attention [3].

Oral administration of HSP65 and β 2-glycoprotein inhibited development of atherosclerosis in experimental models [56].

Induction of immune tolerance through activation of the mucosal immune response, a novel immunomodulating therapy to prevent atherosclerosis, warrants further evaluation [229]. GGA protected the gastric mucosa by inducing HSF1 and HSP70 mRNA [222] and has recently been shown to be cardioprotective by inducing HSP72 [230].

Arimoclomol, a small molecule that induces HSF1, has been shown to increase HSP70 and 90. Interestingly, arimoclomol is currently in phase II/II clinical trials as a treatment for amyotrophic lateral sclerosis [231].

Celastrol is a triterpenoid compound with a retinoid skeleton used in traditional Chinese medicine. This drug potently induces HSF1 and HSP70 expression with antioxidant and anti-inflammatory properties [232].

Statins have many benefits including modulation of the immune system, reduction in apoptosis, and nitricoxide production. Simvastatin and lovastatin induced HSP27 in an osteoblast-like cell line [233].

4. HSPs AS PREDICTORS OF CVD

Although HSPs are present in the sera of healthy individuals and increased in early CVD, their value for predicting atherosclerotic risk has not been comprehensively evaluated.

Pockley *et al.* studied subjects with established hypertension over a 4-year follow-up [95]. Atherosclerosis was assessed by measuring common

carotid artery I-M thickness. Increased I-M thicknesses at follow-up was less prevalent in subjects with increased serum HSP70 (75th percentile) at the time of enrollment. A similar trend was observed for serum HSP60, but was not statistically significant. Anti-HSP was not associated with I-M thickness. The relationship between HSP70 and I-M thickness was independent of age, treatment with atenolol or lacidipine, smoking history, and blood lipids. These findings indicated that circulating HSP70 predicted development of atherosclerosis in established hypertension. An intriguing possibility is that HSP70 protected against or modified atherosclerotic progression [95].

Zhu *et al.* investigated whether combining some risk factors (pathogen burden, increased CRP, and anti-HSP60) enhanced CVD risk. Their results indicated that CVD risk prediction can be improved by combining these three risk factors [234].

Okada *et al.* analyzed the prevalence of the serum IgG antibodies against *H. pylori*-derived HSP60 or its peptide fragments in patients with CVD as compared to those in age- and gender-matched non-CVD patients. Results strongly suggested that IgG against *H. pylori*-derived HSP60 cross-reacted with hHSP60 and that these represented independent CVD markers [235].

Increased anti-HSP65 was associated with subsequent cardiovascular events in a cohort of men with documented coronary artery disease [236]. Xu *et al.* found that increased anti-HSP65 was sustained in persons with severe and progressive carotid disease and was an independent predictor of mortality at 5 years [187]. Gromadzka *et al.* found increased anti-HSP65 and anti-HSP70 after ischemic stroke and suggested that these antibodies were independent risk factors for stroke [237].

Jin *et al.* have reported that anti-HSP70 was a risk factor for ischemic stroke and may be a marker for neuroprotection at early stages [238]. Many mechanisms have been suggested for HSP70 including defense against apoptotic and necrotic cell death in cerebral ischemia [239].

5. IMMUNIZATION AND VACCINATION WITH HSP

Currently, there are several clinical trials evaluating autologous cancer-derived HSP-peptide complexes for treatment of carcinoma and melanoma [240]. Some human vaccines, such as BCG, contain HSP. Although vaccination programs are necessary for maintaining "herd" immunity and the prevention of serious infection, they may lead to increased susceptibility to atherosclerosis. HSP-free vaccines could satisfy the goals of protection from infection and also reduced incidence of CVD [240].

BCG vaccines were first used against human tuberculosis in France in 1921, and have been widely used since the 1950s following recommendations by the World Health Organization [241]. BCG vaccination elicited an immune response to HSP, and in young people with preexisting atherosclerotic lesions, there may be an exacerbation of the disease process [242].

BCG vaccine immunization, which contains HSP, also increased the extent of atherosclerosis in the cholesterol-fed rabbit. Anti-HSP60 in BCG immunized rabbits was correlated with atherosclerotic plaque formation suggesting that the specific immune response to BCG-associated HSP might be proatherogenic [110,145]. Atherosclerotic lesions induced by BCG immunization in the absence of traditional risk factors, such as hyper-cholesterolemia, tend to regress with time, which suggest that the inflammatory response to HSP60 in the absence of other CVD risk factors, is not enough to drive atherogenesis over long period of time (Fig. 2.4) [226].

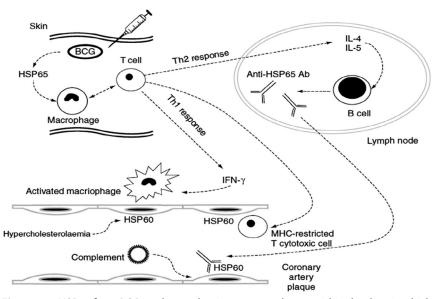


Figure 2.4 HSP65 from BCG is taken up by tissue macrophages within the dermis which present HSP65-derived peptides with class II MHC molecules to either type 1 (Th1) or type 2 (Th2) [186]. Publisher and year of copyright: Elsevier, 2002. Permission for reproduction/adaptation was granted by the copyright holder.

6. EFFECTS OF THERAPEUTIC INTERVENTIONS ON HSP

There have been a few reports on the effects of drugs, herbal medications, and acupuncture on HSP and anti-HSP.

6.1. Herbal medicines

Curcumin extracted from dried rhizomes of the herb *Curcuma longa* (turmeric) and administered (1 g/day) for a period of 30 days had no significant effect on serum anti-HSP27 [243]. Subjects with metabolic syndrome were treated with saffron (100 mg/day) or placebo for 12 weeks and anti-HSP27, 60, 65, and 70 were determined. Saffron was found to decrease anti-HSP27 and 70.

6.2. Statin therapy

Moohebati *et al.* evaluated the effects of statin therapy on serum HSP antibodies in dyslipidemic patients [86]. In this study, 102 subjects were treated with simvastatin (40 mg/day) in a placebo controlled cross-over trial. Simvastatin decreased anti-HSP60, 65, and 70.

7. CONCLUSION

There is good evidence that HSPs are involved in atherogenesis in man and animal models. Although there are a number of options to modulate HSP, it is unclear whether this approach will be beneficial. Tolerization may reduce the immune response to HSP and could be an important factor in mitigating atherosclerotic progression. Several drugs and herbal medicines influence anti-HSP, but their effect is modest and unlikely to provide a viable therapeutic option. While promising, these preliminary studies clearly warrant additional and more comprehensive wellcontrolled prospective studies to clarify the role of HSP and their antibodies in CVD.

REFERENCES

- [1] Y. Chen, T.S. Voegeli, P.P. Liu, E.G. Noble, R.W. Currie, Heat shock paradox and a new role of heat shock proteins and their receptors as anti-inflammation targets, Inflamm. Allergy Drug Targets 6 (2) (2007) 91–100.
- [2] R. Riganò, E. Profumo, B. Buttari, A. Tagliani, L. Petrone, G. D'Amati, et al., Heat shock proteins and autoimmunity in patients with carotid atherosclerosis, Ann. N. Y. Acad. Sci. 1107 (1) (2007) 1–10.

- [3] J. Madrigal-Matute, J.L. Martin-Ventura, L.M. Blanco-Colio, J. Egido, J.-B. Michel, O. Meilhac, Heat-shock proteins in cardiovascular disease, Adv. Clin. Chem. 54 (2011) 1–43.
- [4] R.W. Currie, M. Karmazyn, M. Kloc, K. Mailer, Heat-shock response is associated with enhanced postischemic ventricular recovery, Circ. Res. 63 (3) (1988) 543–549.
- [5] R.A. Williams, Cardiovascular disease in African American women: a health care disparities issue, J. Natl. Med. Assoc. 101 (6) (2009) 536–540.
- [6] R. Ross, The pathogenesis of atherosclerosis: a perspective for the 1990s, Nature 362 (1993) 801.
- [7] H. Donders, J. de Lange, The association between periodontitis and atherosclerosis: the current state of knowledge, J. Cranio-Maxillary Dis. 1 (1) (2012) 17.
- [8] A. Arrigo, W. Welch, Characterization and purification of the small 28,000-dalton mammalian heat shock protein, J. Biol. Chem. 262 (32) (1987) 15359–15369.
- [9] G.A.A. Ferns, Multiple step-variable pathway hypothesis: a reason why predictions fail in atherosclerosis, Med. Hypotheses 71 (6) (2008) 923–926.
- [10] E. Profumo, B. Buttari, L. Saso, R. Capoano, B. Salvati, R. Riganò, T lymphocyte autoreactivity in inflammatory mechanisms regulating atherosclerosis, Scientific World Journal 2012;2012.
- [11] M. Ghayour-Mobarhan, A. Rahsepar, S. Tavallaie, S. Rahsepar, G. Ferns, The potential role of heat shock proteins in cardiovascular disease: evidence from in vitro and in vivo studies, Adv. Clin. Chem. 48 (2009) 27–72.
- [12] C. Lenzi, A. Palazzuoli, N. Giordano, G. Alegente, C. Gonnelli, M.S. Campagna, et al., *H. pylori* infection and systemic antibodies to CagA and heat shock protein 60 in patients with coronary heart disease, World J. Gastroenterol. 12 (48) (2006) 7815.
- [13] Q. Xu, G. Oberhuber, M. Gruschwitz, G. Wick, Immunology of atherosclerosis: cellular composition and major histocompatibility complex class II antigen expression in aortic intima, fatty streaks, and atherosclerotic plaques in young and aged human specimens, Clin. Immunol. Immunopathol. 56 (3) (1990) 344–359.
- [14] P. Libby, Biology of disease. Involvement of the immune system in human atherogenesis: current knowledge and unanswered questions, Lab. Invest. 64 (1991) 5–15.
- [15] C. Napoli, F. D'Armiento, F. Mancini, A. Postiglione, J. Witztum, G. Palumbo, et al., Fatty streak formation occurs in human fetal aortas and is greatly enhanced by maternal hypercholesterolemia. Intimal accumulation of low density lipoprotein and its oxidation precede monocyte recruitment into early atherosclerotic lesions, J. Clin. Investig. 100 (11) (1997) 2680.
- [16] G. Millonig, H. Niederegger, W. Rabl, B. Hochleitner, D. Hoefer, N. Romani, et al., Network of vascular-associated dendritic cells in intima of healthy young individuals, Arterioscler. Thromb. Vasc. Biol. 21 (4) (2001) 503–508.
- [17] R. Kleindienst, Q. Xu, J. Willeit, F. Waldenberger, S. Weimann, G. Wick, Immunology of atherosclerosis. Demonstration of heat shock protein 60 expression and T lymphocytes bearing alpha/beta or gamma/delta receptor in human atherosclerotic lesions, Am. J. Pathol. 142 (6) (1993) 1927.
- [18] G.K. Hansson, Immune mechanisms in atherosclerosis, Arterioscler. Thromb. Vasc. Biol. 21 (12) (2001) 1876–1890.
- [19] G. Millonig, G.T. Malcom, G. Wick, Early inflammatory-immunological lesions in juvenile atherosclerosis from the Pathobiological Determinants of Atherosclerosis in Youth (PDAY)-study, Atherosclerosis 160 (2) (2002) 441–448.
- [20] B. Metzler, M. Mayr, H. Dietrich, M. Singh, E. Wiebe, Q. Xu, et al., Inhibition of arteriosclerosis by T-cell depletion in normocholesterolemic rabbits immunized with heat shock protein 65, Arterioscler. Thromb. Vasc. Biol. 19 (8) (1999) 1905–1911.
- [21] J.T. Grayston, R.A. Kronmal, L.A. Jackson, A.F. Parisi, J.B. Muhlestein, J.D. Cohen, et al., Azithromycin for the secondary prevention of coronary events, N. Engl. J. Med. 352 (16) (2005) 1637–1645.

- [22] A. Van der Wal, P. Das, B.D. Bentz, C. Van der Loos, A. Becker, Atherosclerotic lesions in humans. In situ immunophenotypic analysis suggesting an immune mediated response, Lab. Invest. 61 (2) (1989) 166.
- [23] P.M. Ridker, M.J. Stampfer, N. Rifai, Novel risk factors for systemic atherosclerosis, JAMA 285 (19) (2001) 2481–2485.
- [24] P.M. Ridker, N. Rifai, L. Rose, J.E. Buring, N.R. Cook, Comparison of C-reactive protein and low-density lipoprotein cholesterol levels in the prediction of first cardiovascular events, N. Engl. J. Med. 347 (20) (2002) 1557–1565.
- [25] S. Kiechl, G. Egger, M. Mayr, C.J. Wiedermann, E. Bonora, F. Oberhollenzer, et al., Chronic infections and the risk of carotid atherosclerosis: prospective results from a large population study, Circulation 103 (8) (2001) 1064–1070.
- [26] H. Yu, N. Rifai, High-sensitivity C-reactive protein and atherosclerosis: from theory to therapy, Clin. Biochem. 33 (8) (2000) 601.
- [27] R. Poston, D. Haskard, J. Coucher, N. Gall, R. Johnson-Tidey, Expression of intercellular adhesion molecule-1 in atherosclerotic plaques, Am. J. Pathol. 140 (3) (1992) 665.
- [28] J. Torzewski, D.E. Bowyer, J. Waltenberger, C. Fitzsimmons, Processes in atherogenesis: complement activation, Atherosclerosis 132 (2) (1997) 131–138.
- [29] Z. Prohászka, J. Duba, G. Lakos, E. Kiss, L. Varga, L. Jánoskuti, et al., Antibodies against human heat-shock protein (hsp) 60 and mycobacterial hsp65 differ in their antigen specificity and complement-activating ability, Int. Immunol. 11 (9) (1999) 1363–1370.
- [30] W.J. de Villiers, J.D. Smith, M. Miyata, H.M. Dansky, E. Darley, S. Gordon, Macrophage phenotype in mice deficient in both macrophage-colony—stimulating factor (Op) and apolipoprotein E, Arterioscler. Thromb. Vasc. Biol. 18 (4) (1998) 631–640.
- [31] T. Rajavashisth, J.-H. Qiao, S. Tripathi, J. Tripathi, N. Mishra, M. Hua, et al., Heterozygous osteopetrotic (op) mutation reduces atherosclerosis in LDL receptordeficient mice, J. Clin. Investig. 101 (12) (1998) 2702.
- [32] Z. Mallat, H. Ait-Oufella, A. Tedgui, Regulatory T-cell immunity in atherosclerosis, Trends Cardiovasc. Med. 17 (4) (2007) 113–118.
- [33] P. Szodoray, O. Timar, K. Veres, H. Der, E. Szomjak, G. Lakos, et al., TH1/TH2 imbalance, measured by circulating and intracytoplasmic inflammatory cytokines immunological alterations in acute coronary syndrome and stable coronary artery disease, Scand. J. Immunol. 64 (3) (2006) 336–344.
- [34] C. Heeschen, S. Dimmeler, C.W. Hamm, S. Fichtlscherer, E. Boersma, M.L. Simoons, et al., Serum level of the antiinflammatory cytokine interleukin-10 is an important prognostic determinant in patients with acute coronary syndromes, Circulation 107 (16) (2003) 2109–2114.
- [35] M.S. Elkind, T. Rundek, R.R. Sciacca, R. Ramas, H.J. Chen, B. Boden-Albala, et al., Interleukin-2 levels are associated with carotid artery intima-media thickness, Atherosclerosis 180 (1) (2005) 181–187, Epub 2005/04/13.
- [36] S. Okazaki, S. Furukado, Y. Abe, M. Tanaka, K. Miwa, H. Yamagami, et al., Association of inflammatory markers and carotid intima-media thickness with the risk of cardiovascular events in high-risk patients, Cerebrovasc. Dis. 30 (2) (2010) 180–187, Epub 2010/07/01.
- [37] S.E. Epstein, J. Zhu, A.H. Najafi, M.S. Burnett, Insights into the role of infection in atherogenesis and in plaque rupture, Circulation 119 (24) (2009) 3133–3141.
- [38] M.S. Burnett, C.A. Gaydos, G.E. Madico, S.M. Glad, B. Paigen, T.C. Quinn, et al., Atherosclerosis in apoE knockout mice infected with multiple pathogens, J. Infect. Dis. 183 (2) (2001) 226–231.
- [39] L. Li, E. Messas, E.L. Batista Jr., R.A. Levine, S. Amar, *Porphyromonas gingivalis* infection accelerates the progression of atherosclerosis in a heterozygous apolipoprotein E-deficient murine model, Circulation 105 (7) (2002) 861–867.

- [40] G. Caligiuri, M. Rottenberg, A. Nicoletti, H. Wigzell, G.K. Hansson, *Chlamydia pneumoniae* infection does not induce or modify atherosclerosis in mice, Circulation 103 (23) (2001) 2834–2838.
- [41] K. Ishihara, A. Nabuchi, R. Ito, K. Miyachi, H.K. Kuramitsu, K. Okuda, Correlation between detection rates of periodontopathic bacterial DNA in coronary stenotic artery plaque and in dental plaque samples, J. Clin. Microbiol. 42 (11) (2004) 5437.
- [42] C.-C. Kuo, J.T. Grayston, D.L. Patton, H. Fukushi, L.A. Campbell, A. Shor, Demonstration of *Chlamydia pneumoniae* in atherosclerotic lesions of coronary arteries, J. Infect. Dis. 167 (4) (1993) 841–849.
- [43] B. Chiu, E. Viira, W. Tucker, I. Fong, *Chlamydia pneumoniae*, cytomegalovirus, and herpes simplex virus in atherosclerosis of the carotid artery, Circulation 96 (7) (1997) 2144–2148.
- [44] M.V. Kalayoglu, P. Libby, G.I. Byrne, *Chlamydia pneumoniae* as an emerging risk factor in cardiovascular disease, JAMA 288 (21) (2002) 2724–2731.
- [45] C.M. O'Connor, M.W. Dunne, M.A. Pfeffer, J.B. Muhlestein, L. Yao, S. Gupta, et al., Azithromycin for the secondary prevention of coronary heart disease events, JAMA 290 (11) (2003) 1459–1466.
- [46] B. Cercek, P.K. Shah, M. Noc, D. Zahger, U. Zeymer, S. Matetzky, et al., Effect of short-term treatment with azithromycin on recurrent ischaemic events in patients with acute coronary syndrome in the Azithromycin in Acute Coronary Syndrome (AZACS) trial: a randomised controlled trial, Lancet 361 (9360) (2003) 809–813.
- [47] Q. Xiao, K. Mandal, G. Schett, M. Mayr, G. Wick, F. Oberhollenzer, et al., Association of serum-soluble heat shock protein 60 with carotid atherosclerosis clinical significance determined in a follow-up study, Stroke 36 (12) (2005) 2571–2576.
- [48] C.C. Prelipcean, C. Mihai, P. Gogălniceanu, D. Mitrică, V. Drug, C. Stanciu, Extragastric manifestations of *Helicobacter pylori* infection, Rev. Med.Chir. Soc. Med. Nat. Iași 111 (3) (2007) 575.
- [49] J. Majka, T. Róg, P.C. Konturek, S.J. Konturek, W. Bielański, M. Kowalsky, et al., Influence of chronic *Helicobacter pylori* infection on ischemic cerebral stroke risk factors, Med. Sci. Monit. 8 (10) (2002) CR675.
- [50] S.W. Yusuf, R.M. Mishra, Effect of *Helicobacter pylori* infection on fibrinogen level in elderly patients with ischaemic heart disease, Acta Cardiol. 57 (5) (2002) 317.
- [51] D. Pedicino, A.F. Giglio, V.A. Galiffa, P. Cialdella, F. Trotta, F. Graziani, et al., Infections, immunity and atherosclerosis: pathogenic mechanisms and unsolved questions, Int. J. Cardiol. 3 (2012) 572–583.
- [52] E. Pesonen, I. Paakkari, J. Rapola, Infection-associated intimal thickening in the coronary arteries of children, Atherosclerosis 142 (2) (1999) 425.
- [53] M. Chmiela, M. Kowalewicz-Kulbat, A. Miszczak, M. Wisniewska, T. Rechcinski, K. Kolodziej, et al., A link between *Helicobacter pylori* and/or Chlamydia spp. infections and atherosclerosis, FEMS Immunol. Med. Microbiol. 36 (3) (2006) 187–192.
- [54] C. Espinola-Klein, H.J. Rupprecht, S. Blankenberg, C. Bickel, H. Kopp, G. Rippin, et al., Impact of infectious burden on extent and long-term prognosis of atherosclerosis, Circulation 105 (1) (2002) 15–21.
- [55] M.E. Rosenfeld, Inflammation and atherosclerosis: direct versus indirect mechanisms, Curr. Opin. Pharmacol. 3 (2013) 154–160, Epub 2013/01/30.
- [56] R. Ohashi, H. Mu, Q. Yao, C. Chen, Atherosclerosis: immunopathogenesis and immunotherapy, Med. Sci. Monit. 10 (11) (2004) RA255.
- [57] M. Akerfelt, R.I. Morimoto, L. Sistonen, Heat shock factors: integrators of cell stress, development and lifespan, Nat. Rev. Mol. Cell Biol. 11 (8) (2010) 545–555.
- [58] F. Ritossa, A new puffing pattern induced by temperature shock and DNP in Drosophila, Cell. Mol. Life Sci. 18 (12) (1962) 571–573.

- [59] Q. Xu, B. Metzler, M. Jahangiri, K. Mandal, Molecular chaperones and heat shock proteins in atherosclerosis, Am. J. Physiol. Heart Circ. Physiol. 302 (3) (2012) H506–H514.
- [60] J. Ranford, B. Henderson, Chaperonins in disease: mechanisms, models, and treatments, Mol. Pathol. 55 (4) (2002) 209–213.
- [61] M. Ghayour-Mobarhan, H. Saber, G.A. Ferns, The potential role of heat shock protein 27 in cardiovascular disease, Clin. Chim. Acta 413 (1) (2012) 15–24.
- [62] S. Lindquist, E. Craig, The heat-shock proteins, Annu. Rev. Genet. 22 (1) (1988) 631–677.
- [63] A. Parcellier, S. Gurbuxani, E. Schmitt, E. Solary, C. Garrido, Heat shock proteins, cellular chaperones that modulate mitochondrial cell death pathways, Biochem. Biophys. Res. Commun. 304 (3) (2003) 505–512.
- [64] N. Mounier, A.P. Arrigo, Actin cytoskeleton and small heat shock proteins: how do they interact? Cell Stress Chaperones 7 (2) (2002) 167.
- [65] E.R. Waters, G.J. Lee, E. Vierling, Evolution, structure and function of the small heat shock proteins in plants, J. Exp. Bot. 47 (3) (1996) 325–338.
- [66] R.S. Boston, P.V. Viitanen, E. Vierling, Molecular chaperones and protein folding in plants, Plant Mol. Biol. 32 (1) (1996) 191–222.
- [67] K.J. Mattila, M.S. Nieminen, V.V. Valtonen, V.P. Rasi, Y.A. Kesäniemi, S.L. Syrjälä, et al., Association between dental health and acute myocardial infarction, BMJ 298 (6676) (1989) 779.
- [68] A. Badrichani, D. Stroka, G. Bilbao, D. Curiel, F. Bach, C. Ferran, Bcl-2 and Bcl-X L serve an anti-inflammatory function in endothelial cells through inhibition of NF-kappaB, J. Clin. Investig. 103 (1999) 543–553.
- [69] K. Kato, S. Goto, Y. Inaguma, K. Hasegawa, R. Morishita, T. Asano, Purification and characterization of a 20-kDa protein that is highly homologous to alpha B crystallin, J. Biol. Chem. 269 (21) (1994) 15302–15309.
- [70] I.J. Benjamin, D.R. McMillan, Stress (heat shock) proteins molecular chaperones in cardiovascular biology and disease, Circ. Res. 83 (2) (1998) 117–132.
- [71] O. Kozawa, H. Matsuno, M. Niwa, D. Hatakeyama, Y. Oiso, K. Kato, et al., HSP20, low-molecular-weight heat shock-related protein, acts extracellularly as a regulator of platelet functions: a novel defense mechanism, Life Sci. 72 (2) (2002) 113–124.
- [72] X. Lu, V. Kakkar, The role of heat shock protein (HSP) in atherosclerosis: pathophysiology and clinical opportunities, Curr. Med. Chem. 17 (10) (2010) 957–973.
- [73] D.S. Latchman, Heat shock proteins and cardiac protection, Cardiovasc. Res. 51 (4) (2001) 637–646.
- [74] J.L. Martin-Ventura, V. Nicolas, X. Houard, L.M. Blanco-Colio, A. Leclercq, J. Egido, et al., Biological significance of decreased HSP27 in human atherosclerosis, Arterioscler. Thromb. Vasc. Biol. 26 (6) (2006) 1337–1343.
- [75] L. Santell, N.S. Bartfeld, E.G. Levin, Identification of a protein transiently phosphorylated by activators of endothelial cell function as the heat-shock protein HSP27. A possible role for protein kinase C, Biochem. J. 284 (Pt. 3) (1992) 705.
- [76] J. Martin, E. Hickey, L. Weber, W. Dillmann, R. Mestril, Influence of phosphorylation and oligomerization on the protective role of the small heat shock protein 27 in rat adult cardiomyocytes, Gene Expr. 7 (4–6) (1999) 349.
- [77] A.K. De, K.M. Kodys, B.S. Yeh, C. Miller-Graziano, Exaggerated human monocyte IL-10 concomitant to minimal TNF-alpha induction by heat-shock protein 27 (Hsp27) suggests Hsp27 is primarily an antiinflammatory stimulus, J. Immunol. 165 (7) (2000) 3951–3958, Epub 2000/10/18.
- [78] S. Salinthone, M. Tyagi, W.T. Gerthoffer, Small heat shock proteins in smooth muscle, Pharmacol. Ther. 119 (1) (2008) 44–54.

- [79] B. Dybdahl, S. Slørdahl, A. Waage, P. Kierulf, T. Espevik, A. Sundan, Myocardial ischaemia and the inflammatory response: release of heat shock protein 70 after myocardial infarction, Heart 91 (3) (2005) 299–304.
- [80] S. Shams, S. Shafi, K. Bodman-Smith, P. Williams, S. Mehta, G.A. Ferns, Anti-heat shock protein-27 (Hsp-27) antibody levels in patients with chest pain: association with established cardiovascular risk factors, Clin. Chim. Acta 395 (1) (2008) 42–46.
- [81] Q. Xu, Role of heat shock proteins in atherosclerosis, Arterioscler. Thromb. Vasc. Biol. 22 (10) (2002) 1547–1559.
- [82] D. Chandra, G. Choy, D.G. Tang, Cytosolic accumulation of HSP60 during apoptosis with or without apparent mitochondrial release evidence that its pro-apoptotic or prosurvival functions involve differential interactions with caspase-3, J. Biol. Chem. 282 (43) (2007) 31289–31301.
- [83] S.-C. Kim, J.P. Stice, L. Chen, J.S. Jung, S. Gupta, Y. Wang, et al., Extracellular heat shock protein 60, cardiac myocytes, and apoptosis, Circ. Res. 105 (12) (2009) 1186–1195.
- [84] C.C. Deocaris, S.C. Kaul, R. Wadhwa, On the brotherhood of the mitochondrial chaperones mortalin and heat shock protein 60, Cell Stress Chaperones 11 (2) (2006) 116.
- [85] L.H. Sigal, S. Williams, B. Soltys, R. Gupta, H9724, a monoclonal antibody to *Borrelia burgdorferi's* flagellin, binds to heat shock protein 60 (HSP60) within live neuroblastoma cells: a potential role for HSP60 in peptide hormone signaling and in an autoimmune pathogenesis of the neuropathy of Lyme disease, Cell. Mol. Neurobiol. 21 (5) (2001) 477–495.
- [86] L. Erkkilä, K. Laitinen, K. Haasio, T. Tiirola, M. Jauhiainen, H.A. Lehr, et al., Heat shock protein 60 autoimmunity and early lipid lesions in cholesterol-fed C57BL/ 6JBom mice during *Chlamydia pneumoniae* infection, Atherosclerosis 177 (2) (2004) 321–328.
- [87] A.G. Pockley, R. Wu, C. Lemne, R. Kiessling, U. de Faire, J. Frostegård, Circulating heat shock protein 60 is associated with early cardiovascular disease, Hypertension 36 (2) (2000) 303–307.
- [88] K. Mandal, A. Afzal, S. Brecker, J. Poloniecki, Q. Xu, M. Jahangiri, Association of serum soluble heat shock protein 60 with toll-like receptor 4 polymorphism and severity of coronary artery disease, Heart 92 (5) (2006) 683–685.
- [89] J. Zhu, A.A. Quyyumi, D. Rott, G. Csako, H. Wu, J. Halcox, et al., Antibodies to human heat-shock protein 60 are associated with the presence and severity of coronary artery disease: evidence for an autoimmune component of atherogenesis, Circulation 103 (8) (2001) 1071–1075.
- [90] F. Hoppichler, M. Lechleitner, C. Traweger, G. Schett, A. Dzien, W. Sturm, et al., Changes of serum antibodies to heat-shock protein 65 in coronary heart disease and acute myocardial infarction, Atherosclerosis 126 (2) (1996) 333–338.
- [91] A. Veres, Heart Outcomes Prevention Evaluation (HOPE) Study Investigators, Relationship of anti-60 kDa heat shock protein and anti-cholesterol antibodies to cardiovascular events, Circulation 106 (2002) 2775.
- [92] J. Madrigal-Matute, O. López-Franco, L.M. Blanco-Colio, B. Muñoz-García, P. Ramos-Mozo, L. Ortega, et al., Heat shock protein 90 inhibitors attenuate inflammatory responses in atherosclerosis, Cardiovasc. Res. 86 (2) (2010) 330–337.
- [93] S.D. Morris, D. Cumming, D.S. Latchman, D.M. Yellon, Specific induction of the 70-kD heat stress proteins by the tyrosine kinase inhibitor herbimycin-A protects rat neonatal cardiomyocytes. A new pharmacological route to stress protein expression? J. Clin. Investig. 97 (3) (1996) 706.
- [94] A.D. Johnson, P.A. Berberian, M. Tytell, M.G. Bond, Differential distribution of 70-kD heat shock protein in atherosclerosis. Its potential role in arterial SMC survival, Arterioscler. Thromb. Vasc. Biol. 15 (1) (1995) 27–36.

- [95] A.G. Pockley, A. Georgiades, T. Thulin, U. de Faire, J. Frostegård, Serum heat shock protein 70 levels predict the development of atherosclerosis in subjects with established hypertension, Hypertension 42 (3) (2003) 235–238.
- [96] J.L. Martin-Ventura, A. Leclercq, L.M. Blanco-Colio, J. Egido, P. Rossignol, O. Meilhac, et al., Low plasma levels of HSP70 in patients with carotid atherosclerosis are associated with increased levels of proteolytic markers of neutrophil activation, Atherosclerosis 194 (2) (2007) 334–341.
- [97] I. Herz, R. Rosso, A. Roth, G. Keren, J. George, Serum levels of anti heat shock protein 70 antibodies in patients with stable and unstable angina pectoris, Acute Card. Care 8 (1) (2006) 46–50.
- [98] Q. Xu, S. Kiechl, M. Mayr, B. Metzler, G. Egger, F. Oberhollenzer, et al., Association of serum antibodies to heat-shock protein 65 with carotid atherosclerosis: clinical significance determined in a follow-up study, Circulation 100 (11) (1999) 1169–1174.
- [99] L.-Y. Lin, C.-Y. Lin, T.-C. Su, C.-S. Liau, Angiotensin II-induced apoptosis in human endothelial cells is inhibited by adiponectin through restoration of the association between endothelial nitric oxide synthase and heat shock protein 90, FEBS Lett. 574 (1) (2004) 106–110.
- [100] Y. Shi, D.D. Mosser, R.I. Morimoto, Molecular chaperones as HSF1-specific transcriptional repressors, Genes Dev. 12 (5) (1998) 654–666.
- [101] R. Businaro, E. Profumo, A. Tagliani, B. Buttari, S. Leone, G. D'Amati, et al., Heatshock protein 90: a novel autoantigen in human carotid atherosclerosis, Atherosclerosis 207 (1) (2009) 74–83.
- [102] G. Wick, M. Knoflach, Q. Xu, Autoimmune and inflammatory mechanisms in atherosclerosis, Annu. Rev. Immunol. 22 (2004) 361–403.
- [103] W. Van Eden, G. Wick, S. Albani, I. Cohen, Stress, heat shock proteins, and autoimmunity, Ann. N. Y. Acad. Sci. 1113 (1) (2007) 217–237.
- [104] C. Grundtman, S.B. Kreutmayer, G. Almanzar, M.C. Wick, G. Wick, Heat shock protein 60 and immune inflammatory responses in atherosclerosis, Arterioscler. Thromb. Vasc. Biol. 31 (5) (2011) 960–968.
- [105] P.A. Berberian, W. Myers, M. Tytell, V. Challa, M. Bond, Immunohistochemical localization of heat shock protein-70 in normal-appearing and atherosclerotic specimens of human arteries, Am. J. Pathol. 136 (1) (1990) 71.
- [106] K. Kogure, H. Kato, Altered gene expression in cerebral ischemia, Stroke 24 (12) (1993) 2121–2127.
- [107] R. Shringarpure, T. Grune, J. Mehlhase, K.J. Davies, Ubiquitin conjugation is not required for the degradation of oxidized proteins by proteasome, J. Biol. Chem. 278 (1) (2003) 311–318.
- [108] M. Locke, R.M. Tanguay, Diminished heat shock response in the aged myocardium, Cell Stress Chaperones 1 (4) (1996) 251.
- [109] R.A. Fenton, E.W. Dickson, T.E. Meyer, J.G. Dobson, Aging reduces the cardioprotective effect of ischemic preconditioning in the rat heart, J. Mol. Cell. Cardiol. 32 (7) (2000) 1371–1375.
- [110] D.J. Lamb, G. Ferns, The magnitude of the immune response to heat shock protein-65 following BCG immunisation is associated with the extent of experimental atherosclerosis, Atherosclerosis 165 (2) (2002) 231.
- [111] R.I. Morimoto, Cells in stress: transcriptional activation of heat shock genes, Science 259 (1993) 1409.
- [112] K.K. Donnahoo, B.D. Shames, A.H. Harken, D.R. Meldrum, Review article: the role of tumor necrosis factor in renal ischemia-reperfusion injury, J. Urol. 162 (1) (1999) 196–203.
- [113] S.H. Satyal, D. Chen, S.G. Fox, J.M. Kramer, R.I. Morimoto, Negative regulation of the heat shock transcriptional response by HSBP1, Genes Dev. 12 (13) (1998) 1962–1974.

- [114] M. Ebrahimi, S. Kazemi-Bajestani, M. Ghayour-Mobarhan, G. Ferns, The status of coronary artery disease and its risk factors in Iran: a review, Iran. Red Crescent Med. J. 13 (2011) 615–626.
- [115] G.I. Lancaster, M.A. Febbraio, Exosome-dependent trafficking of HSP70: a novel secretory pathway for cellular stress proteins, J. Biol. Chem. 280 (24) (2005) 23349–23355.
- [116] S. Gupta, A.A. Knowlton, HSP60 trafficking in adult cardiac myocytes: role of the exosomal pathway, Am. J. Physiol. Heart Circ. Physiol. 292 (6) (2007) H3052–H3056.
- [117] K. Rayner, Y.-X. Chen, M. McNulty, T. Simard, X. Zhao, D.J. Wells, et al., Extracellular release of the atheroprotective heat shock protein 27 is mediated by estrogen and competitively inhibits acLDL binding to scavenger receptor-A, Circ. Res. 103 (2) (2008) 133–141.
- [118] K. Heltai, Z. Kis, K. Burian, V. Endresz, A. Veres, E. Ludwig, et al., Elevated antibody levels against *Chlamydia pneumoniae*, human HSP60 and mycobacterial HSP65 are independent risk factors in myocardial infarction and ischaemic heart disease, Atherosclerosis 173 (2) (2004) 337–344.
- [119] T. Huittinen, M. Leinonen, L. Tenkanen, M. Mänttäri, H. Virkkunen, T. Pitkänen, et al., Autoimmunity to human heat shock protein 60, *Chlamydia pneumoniae* infection, and inflammation in predicting coronary risk, Arterioscler. Thromb. Vasc. Biol. 22 (3) (2002) 431–437.
- [120] A.G. Pockley, Heat shock proteins, inflammation, and cardiovascular disease, Circulation 105 (8) (2002) 1012–1017.
- [121] Q. Xu, H. Dietrich, H.J. Steiner, A.M. Gown, B. Schoel, G. Mikuz, et al., Induction of arteriosclerosis in normocholesterolemic rabbits by immunization with heat shock protein 65, Arterioscler. Thromb. Vasc. Biol. 12 (7) (1992) 789–799.
- [122] J. George, Y. Shoenfeld, A. Afek, B. Gilburd, P. Keren, A. Shaish, et al., Enhanced fatty streak formation in C57BL/6J mice by immunization with heat shock protein-65, Arterioscler. Thromb. Vasc. Biol. 19 (3) (1999) 505–510.
- [123] A. Afek, J. George, B. Gilburd, L. Rauova, I. Goldberg, J. Kopolovic, et al., Immunization of low-density lipoprotein receptor deficient (LDL-RD) mice with heat shock protein 65 (HSP-65) promotes early atherosclerosis, J. Autoimmun. 14 (2) (2000) 115–121.
- [124] G. Wick, G. Schett, A. Amberger, R. Kleindienst, Q. Xu, Is atherosclerosis an immunologically mediated disease? Immunol. Today 16 (1) (1995) 27–33.
- [125] P.J. Lindsberg, A.J. Grau, Inflammation and infections as risk factors for ischemic stroke, Stroke 34 (10) (2003) 2518–2532.
- [126] M. Mayr, B. Metzler, S. Kiechl, J. Willeit, G. Schett, Q. Xu, et al., Endothelial cytotoxicity mediated by serum antibodies to heat shock proteins of *Escherichia coli* and *Chlamydia pneumoniae*: immune reactions to heat shock proteins as a possible link between infection and atherosclerosis, Circulation 99 (12) (1999) 1560–1566.
- [127] Q. Xu, G. Luef, S. Weimann, R. Gupta, H. Wolf, G. Wick, Staining of endothelial cells and macrophages in atherosclerotic lesions with human heat-shock proteinreactive antisera, Arterioscler. Thromb. Vasc. Biol. 13 (12) (1993) 1763–1769.
- [128] S.H. Kaufmann, Heat shock proteins and the immune response, Immunol. Today 11 (1990) 129–136.
- [129] J. Lamb, V. Bal, P. Mendez-Samperio, A. Mehiert, A. So, J. Rothbard, et al., Stress proteins may provide a link between the immune response to infection and autoimmunity, Int. Immunol. 1 (2) (1989) 191–196.
- [130] P.M. Res, F. Breedveld, J.A. Van Embden, C. Schaar, W. Van Eden, I. Cohen, et al., Synovial fluid T cell reactivity against 65 kD heat shock protein of mycobacteria in early chronic arthritis, Lancet 332 (8609) (1988) 478–480.

- [131] J. Gaston, P.F. Life, P.J. Jenner, M. Colston, P.A. Bacon, Recognition of a mycobacteria-specific epitope in the 65-kD heat-shock protein by synovial fluidderived T cell clones, J. Exp. Med. 171 (3) (1990) 831–841.
- [132] K.W. Wucherpfennig, J. Newcombe, H. Li, C. Keddy, M.L. Cuzner, D.A. Hafler, Gamma delta T-cell receptor repertoire in acute multiple sclerosis lesions, Proc. Natl. Acad. Sci. USA 89 (10) (1992) 4588–4592.
- [133] C. Georgopoulos, H. McFarland, Heat shock proteins in multiple sclerosis and other autoimmune diseases, Immunol. Today 14 (8) (1993) 373–375.
- [134] D. Child, C. Smith, C. Williams, Heat shock protein and the double insult theory for the development of insulin dependent diabetes, J. R. Soc. Med. 86 (4) (1993) 217.
- [135] R. Tun, M. Smith, S. Lo, G. Rook, P. Lydyard, R. Leslie, Antibodies to heat shock protein 65 kD in type 1 diabetes mellitus, Diabet. Med. 11 (1) (1994) 66–70.
- [136] S.J. Leishman, P.J. Ford, H.L. Do, J.E. Palmer, N.C.K. Heng, M.J. West, et al., Periodontal pathogen load and increased antibody response to heat shock protein 60 in patients with cardiovascular disease, J. Clin. Periodontol. 3 (2012) 923–930.
- [137] J. Van Roon, W. Van Eden, J. Van Roy, F. Lafeber, J. Bijlsma, Stimulation of suppressive T cell responses by human but not bacterial 60-kD heat-shock protein in synovial fluid of patients with rheumatoid arthritis, J. Clin. Investig. 100 (2) (1997) 459.
- [138] S.M. Anderton, R. Van der Zee, B. Prakken, A. Noordzij, W. Van Eden, Activation of T cells recognizing self 60-kD heat shock protein can protect against experimental arthritis, J. Exp. Med. 181 (3) (1995) 943–952.
- [139] V. Pasceri, E.T. Yeh, A tale of two diseases atherosclerosis and rheumatoid arthritis, Circulation 100 (21) (1999) 2124–2126.
- [140] L.E. Hightower, P.T. Guidon, Selective release from cultured mammalian cells of heat-shock (stress) proteins that resemble glia-axon transfer proteins, J. Cell. Physiol. 138 (2) (1989) 257–266.
- [141] S. House, P. Guidon Jr., G. Perdrizet, M. Rewinski, R. Kyriakos, R. Bockman, et al., Effects of heat shock, stannous chloride, and gallium nitrate on the rat inflammatory response, Cell Stress Chaperones 6 (2) (2001) 164.
- [142] M.S. Marber, R. Mestril, S.-H. Chi, M. Sayen, D. Yellon, W. Dillmann, Overexpression of the rat inducible 70-kD heat stress protein in a transgenic mouse increases the resistance of the heart to ischemic injury, J. Clin. Investig. 95 (4) (1995) 1446.
- [143] L.B. Kirby, J. Mondy, C.M. Brophy, Balloon angioplasty induces heat shock protein 70 in human blood vessels, Ann. Vasc. Surg. 13 (5) (1999) 475–479.
- [144] J. George, A. Afek, B. Gilburd, Y. Shoenfeld, D. Harats, Cellular and humoral immune responses to heat shock protein 65 are both involved in promoting fatty-streak formation in LDL-receptor deficient mice, J. Am. Coll. Cardiol. 38 (3) (2001) 900–905.
- [145] D.J. Lamb, L.-J. Eales, G.A. Ferns, Immunization with bacillus Calmette–Guerin vaccine increases aortic atherosclerosis in the cholesterol-fed rabbit, Atherosclerosis 143 (1) (1999) 105–113.
- [146] M. Ghayour-Mobarhan, D.J. Lamb, S. Tavallaie, G.A.A. Ferns, Relationship between plasma cholesterol, von Willebrand factor concentrations, extent of atherosclerosis and antibody titres to heat shock proteins-60, -65 and-70 in cholesterol-fed rabbits, Int. J. Exp. Pathol. 88 (4) (2007) 249–255.
- [147] X. Zhang, X. Wang, H. Zhu, E.G. Kranias, Y. Tang, T. Peng, et al., Hsp20 functions as a novel cardiokine in promoting angiogenesis via activation of VEGFR2, PLoS One 7 (3) (2012) e32765.
- [148] M. Niwa, O. Kozawa, H. Matsuno, K. Kato, T. Uematsu, Small molecular weight heat shock-related protein, HSP20, exhibits an anti-plate activity by inhibiting receptor-mediated calcium influx, Life Sci. 66 (1) (1999) PL7–PL12.

- [149] E.S. Christians, T. Ishiwata, I.J. Benjamin, Small heat shock proteins in redox metabolism: implications for cardiovascular diseases, Int. J. Biochem. Cell Biol. 3 (2012) 1632–1645.
- [150] B.H. Wright, J.M. Corton, A.M. El-Nahas, R.F. Wood, A.G. Pockley, Elevated levels of circulating heat shock protein 70 (Hsp70) in peripheral and renal vascular disease, Heart Vessels 15 (1) (2000) 18–22.
- [151] J. Wysocki, B. Karawajczyk, J. Górski, A. Korzeniowski, Z. Maćkiewicz, G. Kupryszewski, et al., Human heat shock protein 60 (409–424) fragment is recognized by serum antibodies of patients with acute coronary syndromes, Cardiovasc. Pathol. 11 (4) (2002) 238–243.
- [152] G. Novo, F. Cappello, M. Rizzo, G. Fazio, S. Zambuto, E. Tortorici, et al., Hsp60 and heme oxygenase-1 (Hsp32) in acute myocardial infarction, Transl. Res. 157 (5) (2011) 285–292.
- [153] G.-C. Fan, E.G. Kranias, Small heat shock protein 20 (HspB6) in cardiac hypertrophy and failure, J. Mol. Cell. Cardiol. 51 (4) (2011) 574–577.
- [154] G.-C. Fan, X. Ren, J. Qian, Q. Yuan, P. Nicolaou, Y. Wang, et al., Novel cardioprotective role of a small heat-shock protein, Hsp20, against ischemia/reperfusion injury, Circulation 111 (14) (2005) 1792–1799.
- [155] G.-C. Fan, G. Chu, B. Mitton, Q. Song, Q. Yuan, E.G. Kranias, Small heat-shock protein Hsp20 phosphorylation inhibits β-agonist-induced cardiac apoptosis, Circ. Res. 94 (11) (2004) 1474–1482.
- [156] B. Dybdahl, A. Wahba, E. Lien, T.H. Flo, A. Waage, N. Qureshi, et al., Inflammatory response after open heart surgery release of heat-shock protein 70 and signaling through toll-like receptor-4, Circulation 105 (6) (2002) 685–690.
- [157] J. Zhu, A.A. Quyyumi, H. Wu, G. Csako, D. Rott, A. Zalles-Ganley, et al., Increased serum levels of heat shock protein 70 are associated with low risk of coronary artery disease, Arterioscler. Thromb. Vasc. Biol. 23 (6) (2003) 1055–1059.
- [158] A.D. Johnson, P.A. Berberian, M. Tytell, M. Bond, Atherosclerosis alters the localization of HSP70 in human and macaque aortas, Exp. Mol. Pathol. 58 (3) (1993) 155–168.
- [159] S. Genth-Zotz, A.P. Bolger, P.R. Kalra, S. von Haehling, W. Doehner, A.J.S. Coats, et al., Heat shock protein 70 in patients with chronic heart failure: relation to disease severity and survival, Int. J. Cardiol. 96 (3) (2004) 397–401.
- [160] D.R. Ciocca, S.K. Calderwood, Heat shock proteins in cancer: diagnostic, prognostic, predictive, and treatment implications, Cell Stress Chaperones 10 (2) (2005) 86.
- [161] I.A. Yamboliev, J.C. Hedges, J.L.-M. Mutnick, L.P. Adam, W.T. Gerthoffer, Evidence for modulation of smooth muscle force by the p38 MAP kinase/HSP27 pathway, Am. J. Physiol. Heart Circ. Physiol. 278 (6) (2000) H1899–H1907.
- [162] G. Ferns, S. Shams, S. Shafi, Heat shock protein 27: its potential role in vascular disease, Int. J. Exp. Pathol. 87 (4) (2006) 253–274.
- [163] J.L. Martin, R. Mestril, R. Hilal-Dandan, L.L. Brunton, W.H. Dillmann, Small heat shock proteins and protection against ischemic injury in cardiac myocytes, Circulation 96 (12) (1997) 4343–4348.
- [164] R.S. Vander Heide, Increased expression of HSP27 protects canine myocytes from simulated ischemia-reperfusion injury, Am. J. Physiol. Heart Circ. Physiol. 282 (3) (2002) H935–H941.
- [165] G. Kohn, H.R. Wong, K. Bshesh, B. Zhao, N. Vasi, A. Denenberg, et al., Heat shock inhibits tnf-induced ICAM-1 expression in human endothelial cells via I kappa kinase inhibition, Shock 17 (2) (2002) 91–97.
- [166] M. Ghayour-Mobarhan, A. Sahebkar, S.M. Parizadeh, M. Moohebati, S. Tavallaie, S.M. Rezakazemi-Bajestani, et al., Antibody titres to heat shock protein 27 are elevated in patients with acute coronary syndrome, Int. J. Exp. Pathol. 89 (3) (2008) 209–215, Epub 2008/05/08.

- [167] H. Pourghadamyari, M. Moohebati, S.M.R. Parizadeh, H. Falsoleiman, M. Dehghani, A. Fazlinezhad, et al., Serum antibody titers against heat shock protein 27 are associated with the severity of coronary artery disease, Cell Stress Chaperones 16 (3) (2011) 309–316.
- [168] E.W. Peng, D. McCaig, J. Pollock, K. MacArthur, F. Lyall, M.H. Danton, Myocardial expression of heat shock protein 70i protects early postoperative right ventricular function in cyanotic tetralogy of Fallot, J. Thorac. Cardiovasc. Surg. 141 (5) (2011) 1184–1191.
- [169] H.K. Park, E.-C. Park, S.W. Bae, M.Y. Park, S.W. Kim, H.S. Yoo, et al., Expression of heat shock protein 27 in human atherosclerotic plaques and increased plasma level of heat shock protein 27 in patients with acute coronary syndrome, Circulation 114 (9) (2006) 886–893.
- [170] K.A. Buzzard, A.J. Giaccia, M. Killender, R.L. Anderson, Heat shock protein 72 modulates pathways of stress-induced apoptosis, J. Biol. Chem. 273 (27) (1998) 17147–17153.
- [171] C. Garrido, S. Gurbuxani, L. Ravagnan, G. Kroemer, Heat shock proteins: endogenous modulators of apoptotic cell death, Biochem. Biophys. Res. Commun. 286 (3) (2001) 433–442.
- [172] U. Feige, W. Van Eden, Infection, Autoimmunity and Autoimmune Disease. Stress-Inducible Cellular Responses, Springer, Switzerland, 1995, pp. 359–373.
- [173] Z. Prohászka, J. Duba, L. Horvath, A. Csaszar, I. Karadi, A. Szebeni, et al., Comparative study on antibodies to human and bacterial 60 kDa heat shock proteins in a large cohort of patients with coronary heart disease and healthy subjects, Eur. J. Clin. Invest. 31 (4) (2001) 285–292.
- [174] I.R. Cohen, D.B. Young, Autoimmunity, microbial immunity and the immunological homunculus, Immunol. Today 12 (4) (1991) 105–110, Epub 1991/04/01.
- [175] A. Machalinska, M.P. Kawa, W. Marlicz, B. Machalinski, Complement system activation and endothelial dysfunction in patients with age-related macular degeneration (AMD): possible relationship between AMD and atherosclerosis, Acta Ophthalmol. 90 (8) (2012) 695–703, Epub 2011/11/10.
- [176] R. Ross, Atherosclerosis—an inflammatory disease, N. Engl. J. Med. 340 (2) (1999) 115–126, Epub 1999/01/14.
- [177] B.J. Brundel, R.H. Henning, L. Ke, I.C. Van Gelder, H.J. Crijns, H.H. Kampinga, Heat shock protein upregulation protects against pacing-induced myolysis in HL-1 atrial myocytes and in human atrial fibrillation, J. Mol. Cell. Cardiol. 41 (3) (2006) 555–562, Epub 2006/08/01.
- [178] J. Zhu, R.J. Katz, A.A. Quyyumi, D.A. Canos, D. Rott, G. Csako, et al., Association of serum antibodies to heat-shock protein 65 with coronary calcification levels: suggestion of pathogen-triggered autoimmunity in early atherosclerosis, Circulation 109 (1) (2004) 36–41, Epub 2003/12/10.
- [179] M. Mukherjee, C. De Benedictis, D. Jewitt, V.V. Kakkar, Association of antibodies to heat-shock protein-65 with percutaneous transluminal coronary angioplasty and subsequent restenosis, Thromb. Haemost. 75 (2) (1996) 258–260, Epub 1996/02/01.
- [180] A.I. Ibitayo, J. Sladick, S. Tuteja, O. Louis-Jacques, H. Yamada, G. Groblewski, et al., HSP27 in signal transduction and association with contractile proteins in smooth muscle cells, Am. J. Physiol. Gastrointest. Liver Physiol. 277 (2) (1999) G445–G454.
- [181] C. Gisolfi, R. Matthes, K. Kregel, R. Oppliger, Splanchnic sympathetic nerve activity and circulating catecholamines in the hyperthermic rat, J. Appl. Physiol. 70 (4) (1991) 1821–1826.
- [182] Å. Hobbesland, H. Kjuus, D.S. Thelle, Mortality from cardiovascular diseases and sudden death in ferroalloy plants, Scand. J. Work Environ. Health 1997 (1997) 334–341.

- [183] Q. Xu, G. Schett, H. Perschinka, M. Mayr, G. Egger, F. Oberhollenzer, et al., Serum soluble heat shock protein 60 is elevated in subjects with atherosclerosis in a general population, Circulation 102 (1) (2000) 14–20.
- [184] Y.-O. Shin, J.-S. Bae, J.-B. Lee, J.-K. Kim, Y.-J. Kim, C. Kim, et al., Effect of cardiac rehabilitation and statin treatment on anti-HSP antibody titers in patients with coronary artery disease after percutaneous coronary intervention, Int. Heart J. 47 (5) (2006) 671–682.
- [185] F. Belli, A. Testori, L. Rivoltini, M. Maio, G. Andreola, M.R. Sertoli, et al., Vaccination of metastatic melanoma patients with autologous tumor-derived heat shock protein gp96-peptide complexes: clinical and immunologic findings, J. Clin. Oncol. 20 (20) (2002) 4169–4180.
- [186] D.J. Lamb, W. El-Sankary, G. Ferns, Molecular mimicry in atherosclerosis: a role for heat shock proteins in immunisation, Atherosclerosis 167 (2) (2003) 177.
- [187] Q. Xu, G. Wick, J. Willeit, M. Marosi, S. Kiechl, G. Luef, et al., Association of serum antibodies to heat-shock protein 65 with carotid atherosclerosis, Lancet 341 (8840) (1993) 255–259.
- [188] J. Frostegård, C. Lemne, B. Andersson, R. Van der Zee, R. Kiessling, U. de Faire, Association of serum antibodies to heat-shock protein 65 with borderline hypertension, Hypertension 29 (1) (1997) 40–44.
- [189] V. Mazzaferro, J. Coppa, M.G. Carrabba, L. Rivoltini, M. Schiavo, E. Regalia, et al., Vaccination with autologous tumor-derived heat-shock protein gp96 after liver resection for metastatic colorectal cancer, Clin. Cancer Res. 9 (9) (2003) 3235–3245.
- [190] G. Schett, Q. Xu, A. Amberger, R. Van der Zee, H. Recheis, J. Willeit, et al., Autoantibodies against heat shock protein 60 mediate endothelial cytotoxicity, J. Clin. Investig. 96 (6) (1995) 2569.
- [191] S. Arata, S. Hamaguchi, K. Nose, Effects of the overexpression of the small heat shock protein, HSP27, on the sensitivity of human fibroblast cells exposed to oxidative stress, J. Cell. Physiol. 163 (3) (2005) 458–465.
- [192] S. Arata, S. Hamaguchi, K. Nose, Inhibition of colony formation of NIH 3T3 cells by the expression of the small molecular weight heat shock protein HSP27: involvement of its phosphorylation and aggregation at the C-terminal region, J. Cell. Physiol. 170 (1) (1997) 19–26.
- [193] T.D. Pollard, G.G. Borisy, Cellular motility driven by assembly and disassembly of actin filaments, Cell 112 (4) (2003) 453–465.
- [194] M.S. Elkind, Inflammatory mechanisms of stroke, Stroke 41 (10 Suppl. 1) (2010) S3–S8.
- [195] M. Kowalski, P. Konturek, P. Pieniazek, E. Karczewska, A. Kluczka, R. Grove, et al., Prevalence of *Helicobacter pylori* infection in coronary artery disease and effect of its eradication an coronary lumen reduction after percutations coronary angioplasty, Dig. Liver Dis. 33 (3) (2001) 222–229.
- [196] Z. Banecka-Majkutewicz, W. Sawuła, L. Kadziński, A. Węgrzyn, B. Banecki, Homocysteine, heat shock proteins, genistein and vitamins in ischemic stroke—pathogenic and therapeutic implications, Acta Biochim. Pol. 3 (2012) 495–499.
- [197] J.C. Borges, C.H. Ramos, Characterization of nucleotide-induced changes on the quaternary structure of human 70 kDa heat shock protein Hsp70. 1 by analytical ultracentrifugation, BMB Rep. 42 (3) (2009) 166–171.
- [198] Y. Bulut, E. Faure, L. Thomas, H. Karahashi, K.S. Michelsen, O. Equils, et al., Chlamydial heat shock protein 60 activates macrophages and endothelial cells through tolllike receptor 4 and MD2 in a MyD88-dependent pathway, J. Immunol. 168 (3) (2002) 1435–1440.
- [199] D.F.P. Burut, A. Borai, C. Livingstone, G. Ferns, Serum heat shock protein 27 antigen and antibody levels appear to be related to the macrovascular complications associated with insulin resistance: a pilot study, Cell Stress Chaperones 15 (4) (2010) 379–386.

- [200] J.-M. Bruey, C. Ducasse, P. Bonniaud, L. Ravagnan, S.A. Susin, C. Diaz-Latoud, et al., Hsp27 negatively regulates cell death by interacting with cytochrome c, Nat. Cell Biol. 2 (9) (2000) 645–652.
- [201] I.J. Danan, E.R. Rashed, C. Depre, Therapeutic potential of H11 kinase for the ischemic heart, Cardiovasc. Drug Rev. 25 (1) (2007) 14–29.
- [202] A. Figueredo, J. Ibarra, A. Rodriguez, A. Molino, E. la Concha, A. Fernandez-Cruz, et al., Increased serum levels of IgA antibodies to hsp70 protein in patients with diabetes mellitus: their relationship with vascular complications, Clin. Immunol. Immunopathol. 79 (3) (1996) 252–255.
- [203] A.G. Pockley, U. de Faire, R. Kiessling, C. Lemne, T. Thulin, J. Frostegård, Circulating heat shock protein and heat shock protein antibody levels in established hypertension, J. Hypertens. 20 (9) (2002) 1815–1820.
- [204] K. Tanonaka, H. Yoshida, W. Toga, K.-i. Furuhama, S. Takeo, Myocardial heat shock proteins during the development of heart failure, Biochem. Biophys. Res. Commun. 283 (2) (2001) 520–525.
- [205] A. Kruger, S. Petersen, S. Turkseven, Hyperlipidaemia and cardiovascular disease, Circ. Res. 97 (2005) 524–532.
- [206] H. Kervinen, T. Huittinen, O. Vaarala, M. Leinonen, P. Saikku, V. Manninen, et al., Antibodies to human heat shock protein 60, hypertension and dyslipidemia. A study of joint effects on coronary risk, Atherosclerosis 169 (2) (2003) 339–344.
- [207] H.J. Kee, G.H. Eom, H. Joung, S. Shin, J.-R. Kim, Y.K. Cho, et al., Activation of histone deacetylase 2 by inducible heat shock protein 70 in cardiac hypertrophy, Circ. Res. 103 (11) (2008) 1259–1269.
- [208] A.R. Kumarapeli, H. Su, W. Huang, M. Tang, H. Zheng, K.M. Horak, et al., αB-Crystallin suppresses pressure overload cardiac hypertrophy, Circ. Res. 103 (12) (2008) 1473–1482.
- [209] T.M. Vondriska, Y. Wang, A new (heat) shocking player in cardiac hypertrophy, Circ. Res. 103 (11) (2008) 1194–1196.
- [210] C.L. Zhang, T.A. McKinsey, S. Chang, C.L. Antos, J.A. Hill, E.N. Olson, Class II histone deacetylases act as signal-responsive repressors of cardiac hypertrophy, Cell 110 (4) (2002) 479–488.
- [211] J. Nishizawa, A. Nakai, K. Matsuda, M. Komeda, T. Ban, K. Nagata, Reactive oxygen species play an important role in the activation of heat shock factor 1 in ischemicreperfused heart, Circulation 99 (7) (1999) 934–941.
- [212] C. Depre, J.E. Tomlinson, R.K. Kudej, V. Gaussin, E. Thompson, S.-J. Kim, et al., Gene program for cardiac cell survival induced by transient ischemia in conscious pigs, Proc. Natl. Acad. Sci. U.S.A. 98 (16) (2001) 9336–9341.
- [213] J. Qian, X. Ren, X. Wang, P. Zhang, W.K. Jones, J.D. Molkentin, et al., Blockade of Hsp20 phosphorylation exacerbates cardiac ischemia/reperfusion injury by suppressed autophagy and increased cell death, Circ. Res. 105 (12) (2009) 1223–1231.
- [214] Y. Wang, L. Chen, N. Hagiwara, A. Knowlton, Regulation of heat shock protein 60 and 72 expression in the failing heart, J. Mol. Cell. Cardiol. 48 (2) (2010) 360–366.
- [215] M. Imanshahidi, H. Hosseinzadeh, Pharmacological and therapeutic effects of *Berberis vulgaris* and its active constituent, berberine, Phytother. Res. 22 (8) (2008) 999–1012.
- [216] A.G. Pockley, J. Bulmer, B.M. Hanks, B.H. Wright, Identification of human heat shock protein 60 (Hsp60) and anti-Hsp60 antibodies in the peripheral circulation of normal individuals, Cell Stress Chaperones 4 (1) (1999) 29.
- [217] L. Sidorik, R. Kyyamova, V. Bobyk, L. Kapustian, O. Rozhko, O. Vigontina, et al., Molecular chaperone, HSP60, and cytochrome P450 2E1 co-expression in dilated cardiomyopathy, Cell Biol. Int. 29 (1) (2005) 51–55.
- [218] L. Lin, S.-C. Kim, Y. Wang, S. Gupta, B. Davis, S.I. Simon, et al., HSP60 in heart failure: abnormal distribution and role in cardiac myocyte apoptosis, Am. J. Physiol. Heart Circ. Physiol. 293 (4) (2007) H2238–H2247.

- [219] B. Henderson, Integrating the cell stress response: a new view of molecular chaperones as immunological and physiological homeostatic regulators, Cell Biochem. Funct. 28 (1) (2010) 1–14.
- [220] T. Niizeki, Y. Takeishi, T. Watanabe, J. Nitobe, T. Miyashita, T. Miyamoto, et al., Relation of serum heat shock protein 60 level to severity and prognosis in chronic heart failure secondary to ischemic or idiopathic dilated cardiomyopathy, Am. J. Cardiol. 102 (5) (2008) 606–610.
- [221] H. Miller, S. Poon, B. Hibbert, K. Rayner, Y.-X. Chen, E.R. O'Brien, Modulation of estrogen signaling by the novel interaction of heat shock protein 27, a biomarker for atherosclerosis, and estrogen receptor β mechanistic insight into the vascular effects of estrogens, Arterioscler. Thromb. Vasc. Biol. 25 (3) (2005) e10–e14.
- [222] W. Bruening, J. Roy, B. Giasson, D.A. Figlewicz, W.E. Mushynski, H.D. Durham, Up-regulation of protein chaperones preserves viability of cells expressing toxic Cu/ Zn-superoxide dismutase mutants associated with amyotrophic lateral sclerosis, J. Neurochem. 72 (2) (1999) 693–699.
- [223] C. Garrido, J.-M. Bruey, A. Fromentin, A. Hammann, A.P. Arrigo, E. Solary, HSP27 inhibits cytochrome c-dependent activation of procaspase-9, FASEB J. 13 (14) (1999) 2061–2070.
- [224] R.K. Ramanathan, M.J. Egorin, C. Erlichman, S.C. Remick, S.S. Ramalingam, C. Naret, et al., Phase I pharmacokinetic and pharmacodynamic study of 17-dimethylaminoethylamino-17-demethoxygeldanamycin, an inhibitor of heatshock protein 90, in patients with advanced solid tumors, J. Clin. Oncol. 28 (9) (2010) 1520–1526.
- [225] F. Ritossa, Discovery of the heat shock response, Cell Stress Chaperones 1 (2) (1996) 97.
- [226] Q. Xu, R. Kleindienst, G. Schett, W. Waitz, S. Jindal, R.S. Gupta, et al., Regression of arteriosclerotic lesions induced by immunization with heat shock protein 65-containing material in normocholesterolemic, but not hypercholesterolemic, rabbits, Atherosclerosis 123 (1) (1996) 145–155.
- [227] W.H. Dillmann, Small heat shock proteins and protection against injury, Ann. N. Y. Acad. Sci. 874 (1) (1999) 66–68.
- [228] B.J. Brundel, L. Ke, A.-J. Dijkhuis, X. Qi, A. Shiroshita-Takeshita, S. Nattel, et al., Heat shock proteins as molecular targets for intervention in atrial fibrillation, Cardiovasc. Res. 78 (3) (2008) 422–428.
- [229] E. Toubi, Y. Shoenfeld, Predictive and protective autoimmunity in cardiovascular diseases: is vaccination therapy a reality? Lupus 14 (9) (2005) 665–669.
- [230] T. Ooie, N. Takahashi, T. Saikawa, T. Nawata, M. Arikawa, K. Yamanaka, et al., Single oral dose of geranylgeranylacetone induces heat-shock protein 72 and renders protection against ischemia/reperfusion injury in rat heart, Circulation 104 (15) (2001) 1837–1843.
- [231] D. Kieran, B. Kalmar, J.R. Dick, J. Riddoch-Contreras, G. Burnstock, L. Greensmith, Treatment with arimoclomol, a coinducer of heat shock proteins, delays disease progression in ALS mice, Nat. Med. 10 (4) (2004) 402–405.
- [232] A.C. Allison, R. Cacabelos, V.R. Lombardi, X.A. Álvarez, C. Vigo, Celastrol, a potent antioxidant and anti-inflammatory drug, as a possible treatment for Alzheimer's disease, Prog. Neuropsychopharmacol. Biol. Psychiatry 25 (7) (2001) 1341–1357.
- [233] X. Wang, H. Tokuda, D. Hatakeyama, K. Hirade, M. Niwa, H. Ito, et al., Mechanism of simvastatin on induction of heat shock protein in osteoblasts, Arch. Biochem. Biophys. 415 (1) (2003) 6–13.
- [234] A.T. Hirsch, Vascular disease, hypertension, and prevention: "from endothelium to clinical events", J. Am. Coll. Cardiol. 42 (2) (2003) 377–379.

- [235] T. Okada, K. Ayada, S. Usui, K. Yokota, J. Cui, Y. Kawahara, et al., Antibodies against heat shock protein 60 derived from *Helicobacter pylori*: diagnostic implications in cardiovascular disease, J. Autoimmun. 29 (2) (2007) 106–115.
- [236] F. Hoppichler, T. Koch, A. Dzien, G. Gschwandtner, M. Lechleitner, Prognostic value of antibody titre to heat-shock protein 65 on cardiovascular events, Cardiology 94 (4) (2001) 220–223.
- [237] G. Gromadzka, J. Zielińska, D. Ryglewicz, U. Fiszer, A. Członkowska, Elevated levels of anti-heat shock protein antibodies in patients with cerebral ischemia, Cerebrovasc. Dis. 12 (3) (2001) 235–239.
- [238] X. Jin, C. Xiao, R.M. Tanguay, L. Yang, F. Wang, M. Chen, et al., Correlation of lymphocyte heat shock protein 70 levels with neurologic deficits in elderly patients with cerebral infarction, Am. J. Med. 117 (6) (2004) 406–411.
- [239] T. Mehta, J. Greenman, C. Ettelaie, A. Venkatasubramaniam, I. Chetter, P. McCollum, Heat shock proteins in vascular disease—a review, Eur. J. Vasc. Endovasc. Surg. 29 (4) (2005) 395–402.
- [240] A. Hoos, D.L. Levey, Vaccination with heat shock protein–peptide complexes: from basic science to clinical applications, Expert Rev. Vaccines 2 (3) (2003) 369–379.
- [241] B. Sander, U. Skansen-Saphir, O. Damm, L. Håkansson, J. Andersson, U. Andersson, Sequential production of Th1 and Th2 cytokines in response to live bacillus Calmette– Guerin, Immunology 86 (4) (1995) 512.
- [242] H. Stary, Evolution and progression of atherosclerotic lesions in coronary arteries of children and young adults, Arteriosclerosis 9 (1 Suppl.) (1989) I19.
- [243] A. Sahebkar, A. Mohammadi, A. Atabati, S. Rahiman, S. Tavallaie, M. Iranshahi, et al., Curcuminoids modulate pro-oxidant-antioxidant balance but not the immune response to heat shock protein 27 and oxidized LDL in obese individuals, Phytother. Res. 27 (12) (2013) 1883–1888, Epub 2013/03/16.